



# Trap Designs, Colors, and Lures for Emerald Ash Borer Detection

Therese M. Poland\*, Toby R. Petrice and Tina M. Ciaramitaro†

USDA Forest Service, Northern Research Station, Lansing, MI, United States

## OPEN ACCESS

### Edited by:

Davide Rassati,  
University of Padova, Italy

### Reviewed by:

Jonathan David Sweeney,  
Canadian Forest Service, Canada

Kevin J. Dodds,  
United States Forest Service,  
United States

### \*Correspondence:

Therese M. Poland  
therese.poland@usda.gov

### † Present address:

Tina M. Ciaramitaro,  
Auburn University School of Forestry  
and Wildlife Sciences, Auburn, AL,  
United States

### Specialty section:

This article was submitted to  
Forest Disturbance,  
a section of the journal  
Frontiers in Forests and Global  
Change

**Received:** 16 August 2019

**Accepted:** 14 November 2019

**Published:** 05 December 2019

### Citation:

Poland TM, Petrice TR and  
Ciaramitaro TM (2019) Trap Designs,  
Colors, and Lures for Emerald Ash  
Borer Detection.  
Front. For. Glob. Change 2:80.  
doi: 10.3389/ffgc.2019.00080

The emerald ash borer, *Agrilus planipennis*, is the most damaging invasive forest insect pest ever to have invaded North America. It is native to Asia and is established in the United States, Canada, European Russia and Ukraine where it threatens native ash across North America and Europe. We evaluated trap designs, colors, and lures for *A. planipennis* detection at sites with varying infestation levels. Purple or green sticky prism traps and multiple funnel traps hung in the canopy of ash trees and double-decker traps (consisting of two sticky prisms attached to a 3m vertical pole at 3m and 1.8m above ground) had high detection rates even at sites with very low infestation levels. At a low infestation site, females were more attracted to dark purple sticky prism traps hung in the canopy and to Manuka oil and Phoebe oil lures than to light green sticky prism traps or cis-3-hexenol lures; whereas, males were more attracted to light green sticky prism traps in the canopy and cis-3-hexenol lures than to dark purple sticky prism traps or Manuka and Phoebe oil lures. More males and females were captured in double-decker traps with dark green upper prisms and light purple lower prisms, baited with cis-3-hexenol, than in double-decker traps with dark purple upper and lower prisms. Dark green funnel traps and double-decker traps with dark green upper and light purple lower prisms baited with cis-3-hexenol lures captured more females than dark green sticky prism traps hung in the canopy at sites with very low infestation levels. Detection rates were similar among trap types and ranged from 75 to 80% for dark green sticky prism traps, 82.5–100% for dark green funnel traps, and 100% for double-decker traps with dark green upper and light purple lower prisms at sites with very low *A. planipennis* infestations. Cost, ability to reuse the traps, and ease of deployment varies among trap types. These and other factors including trap placement, host density and condition should be considered in selecting traps and designing operational surveys. Future research is needed to determine effective trapping radius, relationship of trap catches to population density, cost benefit of different trap types, and optimal deployment strategy. Greater numbers of *A. planipennis* captured and higher detection rates in cis-3-hexenol-baited double-decker traps with dark green upper prisms and light purple lower prisms and in dark green funnel traps compared to dark green prism traps at sites with very low infestation levels, suggest these trap types would be most effective for operational detection surveys.

**Keywords:** *Agrilus planipennis*, invasive species, early detection, multiple funnel trap, prism trap, double-decker trap, color, cis-3-hexenol

## INTRODUCTION

The emerald ash borer, *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae), a wood-boring beetle native to Asia, is the most damaging invasive forest insect pest to have invaded North America (Herms and McCullough, 2014). It was likely introduced in the mid-1990s (Siegert et al., 2014), but was not discovered until 2002 near Detroit, Michigan (Haack et al., 2002; Poland and McCullough, 2006). This invasive pest is spreading rapidly and threatens to eliminate native ash (*Fraxinus* spp.) as an overstory component in North American forests (Herms and McCullough, 2014). It has also been detected and is spreading in European Russia and Ukraine (Haack et al., 2015; Orlova-Bienkowskaja et al., 2019) and is of great concern to other European countries (Orlova-Bienkowskaja, 2014; Musolin et al., 2017).

Early detection and monitoring are critically important for managing invasive species because management becomes increasingly difficult as populations expand and build to high densities (Brockerhoff et al., 2010). Discovering an invasive species at an early stage of invasion before it becomes widespread allows the rapid implementation of risk mitigation measures, efficient and environmentally sound management decisions, and responses to prevent or slow its spread (Liebhold and Tobin, 2008). Developing optimal detection strategies and tools increases the likelihood for early detection and successful invasive species management programs (Tobin et al., 2014). Effective traps are also essential for monitoring established populations to determine relative density of infestations, spread of invasions and location of hot spots, and to evaluate the effectiveness of management strategies and control tactics (Lyons et al., 2008).

Many factors influence trap efficacy including intrinsic trap characteristics such as color, design, and olfactory attractants, as well as extrinsic conditions such as vertical and horizontal trap placement, environmental surroundings (e.g., host abundance, distribution, and condition, and land use such as ports-of-entry, industrial areas, campgrounds, new developments, or forests and natural areas), and population density (Rassati et al., 2015; Poland and Rassati, 2018). There has been considerable research to develop effective traps and other detection tools including trap trees for early detection of *A. planipennis* (McCullough et al., 2009a,b; Ryall, 2015). Electroretinogram assays have demonstrated both male and female *A. planipennis* adults are sensitive to specific wavelengths of violet and green light whereas only females are sensitive to red wavelengths (Crook et al., 2009). Males also orient visually to mates and land on dead pinned females placed on the trunks or leaves of ash trees (Rodríguez-Saona et al., 2007; Lelito et al., 2008) or on artificial decoys (Domingue et al., 2015). In addition to visual cues, *A. planipennis* adults use long- and short-range olfactory cues to locate hosts and mates. Ash volatiles, including bark sesquiterpenes and green leaf volatiles, elicit antennal responses by *A. planipennis* adults (Rodríguez-Saona et al., 2006; Crook et al., 2008). In field studies, adults were attracted to traps baited with the leaf alcohol *cis*-3-hexenol found in the volatiles from ash leaves (Rodríguez-Saona et al., 2006; de Groot et al., 2008; Grant et al., 2010) and natural tree oils including

Manuka oil [from the New Zealand tea tree, *Leptospermum scoparium* J. R. and C. Forst (Myrtaceae)] and Phoebe oil [from the Brazilian walnut tree, *Phoebe porosa* Mez. (Lauraceae)] which contain sesquiterpenes found in ash bark volatiles (Crook et al., 2008; de Groot et al., 2008; Crook and Mastro, 2010; Grant et al., 2010, 2011; Poland et al., 2011). Females produce a combination of contact pheromones that change with reproductive maturity (Lelito et al., 2009; Silk et al., 2009) as well as a short-range volatile pheromone *cis*-3-lactone which has been shown to increase attraction to traps baited with *cis*-3-hexenol (Silk et al., 2011, 2019; Ryall et al., 2012; Ryall, 2015).

Several different trap designs have been developed and evaluated for capture of *A. planipennis*. Early trap designs included box-shaped traps and cross-vane panel traps of different colors (Francese et al., 2008). As our understanding of *A. planipennis* behavior and responses to colors and volatiles advanced (Lelito et al., 2007; Rodríguez-Saona et al., 2007; Crook et al., 2009, 2012; Francese et al., 2010a) trap colors were optimized to specific shades of green or purple and new trap designs were developed. Three-sided prism traps presented a wider silhouette than box or cross-vane traps for the same amount of material and the flat surface resulted in higher reflectance of colors compared to cross-vane traps and a broader sticky area for insect capture (Francese et al., 2008). Sticky cards or leaves with pinned dead *A. planipennis* adults (Lelito et al., 2008; Domingue et al., 2013; Petrice et al., 2013) and branch traps with artificial beetle decoys (Domingue et al., 2015) have been tested to exploit the observation that male *A. planipennis* respond visually to females and land on them in the canopies of ash trees (Lelito et al., 2008). Green and/or purple double-decker traps were designed to incorporate multiple attractive stimuli of ash trees. They are composed of a 10 cm diameter, 3 m tall PVC pole with an upper prism trap attached to the top and a lower prism attached 60 cm below which are baited separately with leaf and bark volatiles, respectively, to represent the tree canopy and bole (Poland et al., 2011). More recently, green or purple multiple-funnel traps with different slippery coatings that can be re-used for multiple years have been tested as an alternative to sticky traps that must be discarded (Francese et al., 2011, 2019; Crook et al., 2014).

In field trapping experiments, green ( $\lambda = 530\text{--}540$  nm wavelengths) and purple (with peak wavelengths at  $\lambda = 430, 600,$  and  $670$  nm) traps were found to be highly attractive to *A. planipennis* (Crook et al., 2009, 2012; Francese et al., 2010a,b, 2013a). Decreasing the reflectance of green traps from 67% to 49% resulted in a darker shade of green and significantly increased the number of adults captured (Francese et al., 2010a), while altering the shade of purple (430 nm 20% reflectance, 600 nm 6% reflectance, and 670 nm 13.5% reflectance) to a lighter shade (420 nm 21.7% reflectance and 670 nm 13.6% reflectance), referred to as Sabic purple, also increased attraction (Francese et al., 2010a, 2011, 2013a). Purple traps were found to be more attractive to females than to males (Crook et al., 2009; Francese et al., 2010b, 2013a).

Trap placement influences *A. planipennis* captures, most likely due to male and female behaviors. In China and North America, *A. planipennis* adults are more active in the afternoon in sunny locations and prefer to attack trees grown in full

sun rather than in the shade (Yu, 1992; McCullough et al., 2009a,b). Males tend to hover around the canopies of ash trees before landing on females to mate or on leaves to feed (Rodriguez-Saona et al., 2007; Lelito et al., 2008). Ryall et al. (2013) captured more *A. planipennis* when green sticky prism traps were placed in the south or sun-exposed aspect of tree canopies compared to shaded aspects. Trap height may also affect capture rates. For instance, Crook et al. (2008) found that more *A. planipennis* were captured on purple prism traps placed high in the canopy compared with similar traps placed below the canopy.

Tree condition and *A. planipennis* infestation levels can also influence trap captures. At high *A. planipennis* densities, heavily infested trees may emit stress-induced volatiles (Rodriguez-Saona et al., 2006) which may enhance attraction to traps hung in infested trees or compete with traps hung in nearby uninfested trees. As canopy dieback progresses in heavy infestations, light availability increases and hyperspectral signatures of trees are altered (Bartels et al., 2008), potentially affecting beetle response to trap color or placement. The number of *A. planipennis* captured in traps hung in the ash canopy was positively correlated with the infestation level of the tree in which they were hung (Poland and McCullough, 2014).

The US Department of Agriculture, Animal and Plant Health Inspection Service, Plant Protection and Quarantine (USDA APHIS-PPQ) Emerald Ash Borer Cooperative Program conducts a multi-state emerald ash borer survey program that relies on traps. The program has evolved with advances in detection techniques from visual surveys in 2002, to girdled trap trees in 2005, and implementation of glue-coated dark purple prism traps baited with Manuka oil in 2008 (USDA-APHIS-PPQ, 2018). Based on positive detection results and feedback, the APHIS survey continues to rely primarily on purple prism traps which are currently baited with *cis*-3-hexenol lures, with 12,000 set out in 2018 in a buffer zone along the leading edge of the infestation. Dark green multiple funnel traps baited with *cis*-3-hexenol are also recommended as an alternative and used in some locations (USDA-APHIS-PPQ, 2018; Francese et al., 2019). In Canada, visual and branch sampling surveys along with green prism traps hung in the canopy of ash trees and baited with *cis*-3-hexenol and *cis*-3-lactone are the recommended tools for *A. planipennis* detection surveys (CFIA, 2019; Silk et al., 2019). Regulatory agencies review and update survey protocols and guidelines annually based on recommendations from science advisory panels, logistical and economic considerations, and consistency of methods and messaging from year to year for monitoring, training, and public outreach purposes.

Although many studies have compared various trap types, colors, and lures (Marshall et al., 2009, 2010b; McCullough et al., 2011; Poland et al., 2011; Crook et al., 2012, 2014; Fierke et al., 2013; Francese et al., 2013a; Poland and McCullough, 2014), results have been variable (Ryall, 2015). Several studies have focused on optimizing the color of sticky prism traps or multiple funnel traps (Crook et al., 2009, 2012; Francese et al., 2010a,b, 2013a), while other studies have compared sticky prism traps baited with different lures (Crook et al.,

2008; de Groot et al., 2008; Grant et al., 2010, 2011; Silk et al., 2011; Ryall et al., 2013; Ryall, 2015). Most studies have been conducted at sites with moderate to high *A. planipennis* infestation levels which are not representative of newly invaded areas with low infestation levels where detection surveys would occur (Burr et al., 2018). Few studies have compared multiple trap types of different colors and baited with different lures to determine optimal recommendations for operational detection programs. Also, optimal colors have not been previously tested for double-decker traps and they have not been compared to multiple trap types including prism traps, and multiple funnel traps.

Our objective was to evaluate the most promising trap designs, colors, and lures for *A. planipennis* detection in order to improve operational survey programs. Using field sites with varying levels of *A. planipennis* infestation, we compared multiple combinations of lures, colors, and trap types including sticky prism and multiple funnel traps hung in the ash canopy and double-decker traps at ground level.

## METHODS

Four trapping experiments were conducted from 2010 to 2015 to compare efficacy of the most promising trap designs for capturing *A. planipennis* (Table 1). Experiments were conducted at field sites in southern Michigan differing in infestation levels of *A. planipennis* based on visible signs and symptoms such as woodpecker attacks, epicormic shoots, and percentage of crown dieback. In 2010, field sites included the MSU Kellogg Experimental Forest, Augusta, Kalamazoo County; Legg Park, Okemos, Ingham County; and Fox Memorial Park, Pottsville, Eaton County. The site at the Kellogg Experimental Forest consisted of a small stand of green ash, *Fraxinus pennsylvanica* Marshall, trees with a moderate infestation of *A. planipennis*. Some trees had evidence of *A. planipennis* infestation including woodpecker feeding holes, epicormic shoots, and about 25–30% crown dieback. Legg Park consisted of a natural area along the Red Cedar River with open fields surrounded by a mixed hardwood forest with a major component of mature green ash. Fox Memorial Park consisted of an open area with a small lake surrounded by mixed hardwoods including a major component of green ash. At Legg Park and Fox Memorial Park, *A. planipennis* infestations were very high with many heavily infested trees having abundant woodpecker attacks, epicormic shoots and >30% crown dieback. In 2014, field sites included Legg Park, Fox Memorial Park, and a private woodlot near Ithaca, Gratiot County, MI. Infestations at Legg Park and Fox Memorial Park had passed their peak and *A. planipennis* populations had declined to very low levels, with few surviving mature ash trees and many small healthy saplings. The private woodlot near Ithaca included a major component of green ash, many of which had symptoms of infestation including woodpecker attacks, epicormic shoots, and 35–40% crown dieback. The population of *A. planipennis* was high and the infestation level was nearing its peak. In 2015, the experiment was conducted at Legg Park where

**TABLE 1** | Field sites and treatments for *A. planipennis* trapping experiments in southern Michigan, 2010–2015.

Experiment number (Year)	Field site	<i>A. planipennis</i> infestation level	Number of blocks	Treatments
1 (2010)	Kellogg Forest	Moderate	2	1) Dark purple and light green prisms unbaited Control
	Legg Park	High	8	2) Dark purple and light green prisms Hex
	Fox Memorial	Very high	7	3) Dark purple and light green prisms Man/Phoebe 4) Dark purple and light green prisms Hex + Man/Phoebe
2 (2014)	Legg Park	Very low	10	1) Dark purple DD HexHex 2) Dark purple DD HexMan 3) Light purple DD HexMan 4) Green/purple DD HexHex 5) Green/purple DD HexMan
3 (2014)	Fox Memorial	Very low	5	1) Dark green funnel Hex
	Ithaca	High	5	2) Dark green prism Hex 3) Green/purple DD HexHex 4) Dark purple DD HexMan
4 (2015)	Legg Park	Very low	8	1) Dark green funnel Hex 2) Dark green prism Hex 3) Green/purple DD HexHex

Trap types included dark or light green or purple prism traps hung in the canopy (prism), dark green multiple funnel traps hung in the canopy (funnel), or double-decker traps composed of two prisms mounted at 3 m and 1.8 m on a vertical 3 m pole (DD), with dark purple for both prisms, light purple for both prisms, or dark green for the upper and light purple for the lower prism (Green/purple). Lures tested included two *cis*-3-hexenol bubble caps released at a combined rate of 7.4 mg/day (Hex), a pouch releasing Manuka and Phoebe oil at 50 mg/day (Man/Phoebe), or a pouch releasing Manuka oil at 50 mg/day (Man). In experiment 1, two prism traps of different colors were hung in each tree and baited with the same lure treatment.

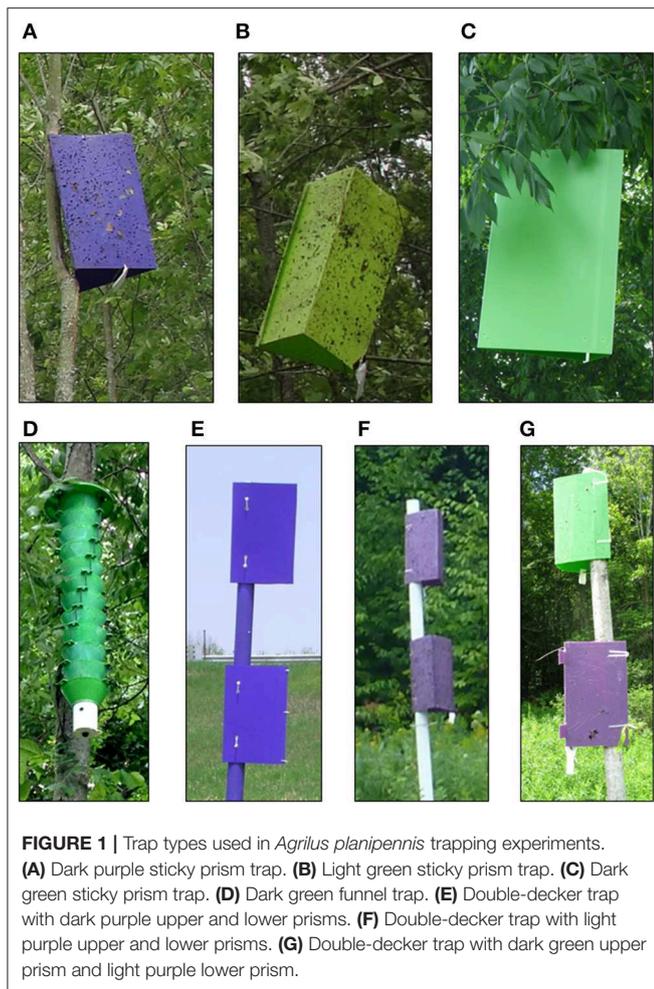
the *A. planipennis* infestation had collapsed and populations remained very low.

We tested three different trap types in various colors. Sticky prism traps consisted of 3-sided corrugated plastic prisms (60 × 40 cm on each side) coated with Pestick insect trapping glue (Hummert International, Earth City, MO). They were hung in the upper half of the canopies of ash trees along the edge of a stand approximately 6–10 m above ground. Traps were hung from rope installed over the end of a branch extending into the opening at the edge of the stand using a Big Shot Line Launcher (Sherrill Tree, Greensboro, NC). Double-decker traps consisted of a 3-m tall PVC pipe (10 cm diameter) set over a T-post in the ground. Two sticky prism traps coated with Pestick were attached to the PVC pipe: an upper prism at the top and a lower prism 60 cm below the upper prism (Poland et al., 2011). Double-decker traps were placed about 5–10 m from the edge of a stand. Multiple funnel traps were standard 12-unit traps (Synergy Semiochemical, Delta, B.C.) and were coated with 10% Fluon, a slippery fluoropolymer (AGC Chemicals Americas, Inc., Exton, PA) which has been shown to enhance captures of woodboring beetles (Graham and Poland, 2012; Allison, 2016). We tested four colors: (1) dark purple ( $\lambda = 430$  nm 20% reflectance, 600 nm 6% reflectance, and 670 nm 13.5% reflectance, Harbor Sales Inc., Suddlersville, MD); (2) light purple ( $\lambda = 420$  nm 21.7% reflectance and 670 nm 13.6% reflectance, “Sabic purple,” Great Lakes Integrated Pest Management, Vestaburg, MI); (3) light green ( $\lambda = 540$  nm, peak reflectance 64%, Great Lakes Integrated Pest Management, Vestaburg, MI); and dark green ( $\lambda = 540$  nm, peak reflectance 49%, Great Lakes Integrated Pest Management, Vestaburg, MI). The light shade of purple and dark shade of green were found to be more attractive to *A. planipennis* than the

original dark purple and light green (Francese et al., 2010a, 2011, 2013a). Prisms of double-decker traps were either both the same color (e.g., “dark purple,” or “light purple”) or different colors for the upper and lower prisms (e.g., dark green upper and light purple lower, hereinafter “green/purple”).

Traps were baited with either two *cis*-3-hexenol bubble caps (hereinafter “Hex”) with a combined release rate of 7.4 mg/day (previously available from the former Contech Enterprises Inc., Delta, B.C., currently similar lures available from Scentry Biologicals, Inc., Billings, MT; Synergy Semiochemicals, Delta, B.C.; Sylvar Technologies, Inc., Fredericton, NB), one pouch with an 80:20 blend of Manuka and Phoebe oil (hereinafter “Man/Phoebe”) released at 50 mg/day (Synergy Semiochemicals Inc., Delta, B.C.), one Manuka oil pouch (hereinafter “Man”) released at 50 mg/day (Synergy Semiochemicals Inc., Delta, B.C.) or different combinations. The upper and lower prisms of double-decker traps were baited separately to provide a point source of attraction for each trapping surface area, with either Hex lures on both prisms, or a Hex lure on the upper panel representing leaf volatiles in the canopy and a Man lure on the lower panel representing bark volatiles from the bole of trees.

Experiment 1 was conducted 12 May–4 August 2010 and compared dark purple and light green sticky prism traps. The experiment used a multi-location randomized complete block factorial design with 4 lure and 2 color treatments and 17 blocks. Two blocks were set up at the Kellogg Experimental Forest with a moderate *A. planipennis* infestation, 8 blocks at Legg Park with a high infestation, and 7 blocks at Fox Memorial Park with a very high infestation. Live ash trees were selected in which to hang the traps. Trees within a block were separated by at least 10 m and blocks were separated by at least 20 m. In each tree,



one dark purple (Figure 1A) and one light green (Figure 1B) sticky prism trap were randomly assigned to separate branches at least 3 m apart. Both traps in the same tree were baited with the same lure treatment. Each block consisted of four trees, each containing a purple and a green trap baited with one of four randomly assigned lure treatments: (1) Unbaited control; (2) Hex; (3) Man/Phoebe; or (4) Both Hex and Man/Pheobe. Lures were replaced on 12 to 13 July at which time infestation level of each tree containing traps was estimated. Infestation level was ranked as “1” if the tree was lightly infested with few visible symptoms and < 10% dieback, “2” if the tree was moderately infested with a few woodpecker feeding holes and epicormic shoots and < 35% dieback, and “3” if the tree was heavily infested and had abundant woodpecker feeding holes, epicormic shoots, and > 35% dieback.

Experiment 2 was conducted 12 June–11 August 2014 at Legg Park with a very low *A. planipennis* population after the first wave of infestation had declined. The experiment compared double-decker traps with five different color and lure combinations including the standard dark purple used for the APHIS survey program and previous studies with double-decker traps (Poland et al., 2011) as well as newer shades of dark green and light purple

that were found to enhance attraction of *A. planipennis* (Francese et al., 2010a, 2013a). There were 10 blocks of 5 treatments laid out in a randomized complete block design with at least 10 m between traps and 25 m between blocks: (1) Dark purple baited with Hex on both prisms (Dark Purple HexHex; Figure 1E); (2) Dark purple baited with Hex on the upper and Man on the lower prism (Dark Purple HexMan); (3) Light purple baited with Hex on both prisms (Light Purple HexHex; Figure 1F); (4) Green/purple baited with Hex on both prisms (Green/Purple HexHex; Figure 1G); and 5) Green/purple baited with Hex on the upper and Man on the lower prism (Green/Purple HexMan).

Experiment 3 was conducted 12 June–3 September 2014 and compared different trap types at two different sites, a private woodlot near Ithaca, MI with a high *A. planipennis* infestation level, and Fox Memorial Park with a very low population level after the initial outbreak had declined. The experiment had 10 blocks of 4 treatments, with 5 blocks at each site which were analyzed separately. The trap designs and colors tested included the most attractive double-decker trap from Experiment 2, the original dark purple double-decker trap design, and the funnel and prism traps in the newer dark green that was reported to enhance attraction of *A. planipennis* (Crook et al., 2012; Francese et al., 2013a,b). Treatments included: (1) Dark green multiple funnel trap baited with Hex (Green Funnel; Figure 1D); (2) Dark green sticky prism trap baited with Hex (Green Prism; Figure 1C); (3) Green/purple double-decker trap baited with Hex on both prisms (Green/purple HexHex); and (4) Dark purple double-decker trap baited with Hex on the upper and Man on the lower prism (Dark purple HexMan).

Experiment 4 was conducted 9 June–30 July 2015 and compared different trap types at Legg Park with a very low *A. planipennis* population level after the initial infestation had declined. The trap types compared were similar to Experiment 3 except that the dark purple double-decker trap which captured fewer *A. planipennis* than the Green/Purple double-decker trap was not included. The experiment included 8 blocks of 3 treatments: (1) Green funnel baited with Hex (Green Funnel); (2) Dark green sticky prism baited with Hex (Green Prism); and (3) Green/purple double-decker trap baited with Hex on both prisms (Green/Purple HexHex).

For all experiments, captured *A. planipennis* were collected every 2 weeks and soaked in Histoclear II (National Diagnostics, Atlanta, GA) to remove Pestick, except for those captured in multiple funnel traps. All *A. planipennis* were sexed and the total number captured of each sex for the entire season was summed for each trap. All analyses were conducted using the SAS 9.4 for Windows statistical package (SAS Institute, 2012) with an  $\alpha$  level of 0.05. For each experiment, data for the number of *A. planipennis* captured were analyzed by a generalized linear mixed model (PROC GLIMMIX) with separate models analyzed for males, females, and totals of both sexes. For experiment 1, color, treatment, and the interaction of color within treatment were tested as fixed effects. The level of infestation of the tree in which traps were hung and interaction between infestation level and treatment were tested when running through the steps of ANCOVA. Since the interaction term was not significant, only infestation level was included as a covariate in the model.

Site, block within site, and the interaction between color and treatment within site were included as random effects in the model. The Kenward Rogers denominator degrees of freedom adjustment method was used as an option in all models. Degrees of freedom were rounded to the nearest whole number for ease of reporting in the results section. The response distribution was lognormal with the identity link function. Differences among treatments and colors were tested with the Tukey-Kramer means comparison procedure. Separate models were run for each site with color, treatment, and color within treatment as fixed effects, block as a random effect, and infestation level of the tree in which traps were hung as a covariate. For experiments 2, 3, and 4, treatment was tested as a fixed effect and block as a random effect. The response distribution was gamma with the log link function. In all models the distribution and link function that had the best residuals in terms of meeting normality via the Shapiro-Wilks test and homogeneity of variance via the Levene's test were retained. For all experiments, the detection rates for each treatment, or percentage of traps that captured at least one beetle were reported.

## RESULTS

In experiment 1, we captured 14,080 *A. planipennis* adults on prism traps hung in the canopy. Both trap color and lure treatment, but not their interaction, had significant effects on the number of *A. planipennis* captured, but responses varied for males and females, with females generally being more attracted to dark purple traps baited with Manuka and Phoebe oil and males to light green traps baited with *cis*-3-hexenol (Table 2). For all sites combined, significantly more females were captured in dark purple sticky prism traps than in light green sticky prism traps hung in the canopy ( $F = 3.81$ ;  $P = 0.05$ ;  $df = 1, 22$ ) and in traps baited with Manuka and Phoebe oil alone or combined with *cis*-3-hexenol than in unbaited control traps ( $F = 3.03$ ;  $P = 0.03$ ; Table 2;  $df = 3, 10$ ). The number of females captured in traps baited with *cis*-3-hexenol alone was intermediate (Table 2). In contrast, significantly more males were captured in traps baited with *cis*-3-hexenol alone or combined with Manuka and Phoebe oil, than in unbaited control traps ( $F = 7.67$ ;  $P = 0.005$ ; Table 2;  $df = 3, 10$ ). The number of males captured in traps baited with Manuka and Phoebe oil alone was intermediate (Table 2). Detection rates (percentage of traps that captured at least one *A. planipennis*) were similar for sticky prism traps of either color and all lure treatments, with detection rates ranging from 97% to 100%. The covariate, infestation level of the tree in which traps were hung, had a significant effect on the number of *A. planipennis* captured for both sexes, but there was no significant interaction with treatment. Significantly more *A. planipennis* were captured in traps hung in trees with heavy infestation levels ( $134.9 \pm 18$ ) than in trees with moderate ( $93.8 \pm 10$ ) or light infestation levels ( $57.9 \pm 13$ ) ( $F = 9.64$ ;  $P = 0.003$ ;  $df = 1, 67$ ).

The pattern of responses for males and females to different treatments and colors were similar at all sites; however, differences in responses to different treatments and colors tended to be more pronounced at the site with the lowest population

density (Table 2). We captured an average ( $\pm$ SE) of  $66.3 \pm 16$  beetles per trap at the Kellogg Experimental Forest site that had a moderate *A. planipennis* population,  $76.4 \pm 9$  beetles per trap at Legg Park with a high population, and  $145.1 \pm 16$  per trap at Fox Memorial Park with a very high population. At the Kellogg Experimental Forest with the lowest population density, significantly more females were captured in traps baited with Manuka and Phoebe oil ( $31.25 \pm 2.5$ ) than in traps baited with *cis*-3-hexenol ( $9.0 \pm 2.4$ ), unbaited control traps ( $9.5 \pm 1.0$ ) or both lures ( $13.3 \pm 2.9$ ) combined ( $F = 12.3$ ;  $df = 3, 11$ ;  $P = 0.008$ ) and in dark purple ( $18.1 \pm 2.8$ ) than in light green ( $13.4 \pm 2.4$ ) sticky prism traps hung in the canopy ( $F = 4.6$ ;  $df = 1, 11$ ;  $P = 0.05$ ). More males were captured in traps baited with *cis*-3-hexenol alone ( $78.8 \pm 36.0$ ) or combined with Manuka and Phoebe oil ( $82.0 \pm 51.4$ ) than in unbaited control traps ( $14.5 \pm 6.1$ ) or traps baited with Manuka and Phoebe oil alone ( $27.3 \pm 4.2$ ;  $F = 4.17$ ;  $df = 3, 11$ ;  $P = 0.03$ ) and in light green ( $77.1 \pm 29.5$ ) than in dark purple ( $24.1 \pm 6.5$ ) sticky prism traps hung in the canopy ( $F = 6.12$ ;  $df = 1, 11$ ;  $P = 0.03$ ). Because significantly more males were captured than females, overall significantly more *A. planipennis* of both sexes combined were captured in light green ( $95.3 \pm 29$ ) than in dark purple ( $37.5 \pm 6.5$ ) sticky prism traps hung in the canopy ( $F = 5.89$ ;  $P = 0.03$ ;  $df = 1, 11$ ).

We captured 494 *A. planipennis* adults in experiment 2 at Legg Park, a site with very low infestation levels in 2014 (Table 3). The color of sticky prisms on the double-decker traps significantly affected catches but lure treatment did not (Table 3). When traps were baited with Hex, the green/purple double-decker traps captured significantly more females, males, and total *A. planipennis* than did dark purple double-decker traps, while light purple double-decker traps had intermediate catches (Table 3). Similarly, when traps were baited with Hex on the upper and Man on the lower prism, significantly more males and total *A. planipennis* were captured on green/purple than on dark purple double-decker traps, whereas, female trap catch did not differ among trap color combinations. Within the same trap color combination, lure treatment had no effect on *A. planipennis* catches. The detection rate of green/purple double-decker traps was substantially higher (100%) than detection rates of dark purple double-decker traps (60%, 70%), regardless of lure.

We captured 1314 *A. planipennis* adults in experiment 3 with 213 at Fox Memorial Park and 1101 at the private woodlot near Ithaca. More *A. planipennis* were captured per trap near Ithaca ( $55.1 \pm 11$ ) with a high infestation level than at Fox Memorial Park ( $10.1 \pm 2$ ) where the *A. planipennis* infestation was past its peak and populations had declined to very low levels ( $F = 31.5$ ;  $df = 1, 8$ ;  $P = 0.0006$ ). At the site near Ithaca with a high *A. planipennis* population, significantly more males and females were captured in dark green funnel and dark green sticky prism traps hung in the canopy than in dark purple double-decker traps. The number of males and females captured in green/purple double-decker traps was intermediate (Table 4). All traps had 100% detection rates. At Fox Memorial Park with a very low infestation level, significantly more females were captured in dark green funnel traps and in green/purple double-decker than in

**TABLE 2** | Mean number ( $\pm$  SE) of *A. planipennis* captured and detection rate (percentage of traps that captured at least one beetle) in Experiment 1 that compared green or purple sticky prism traps baited with a blend of Manuka and Phoebe oil (50 mg/day), *cis*-3-hexenol (7.4 mg/day), or both.

Trap color and lure	Number of traps	Mean number of EAB captured $\pm$ SE			Detection rate (%)
		Females	Males	Total	
<b>Both colors</b>					
Control	34	32.2 $\pm$ 5.8 b	31.3 $\pm$ 5.0 b	64.4 $\pm$ 10.9 b	97 $\pm$ 3
Manuka/Phoebe Oil	34	46.8 $\pm$ 6.6 a	42.8 $\pm$ 6.9 ab	94.8 $\pm$ 15.1 ab	97 $\pm$ 3
<i>cis</i> -3-Hexenol	34	38.5 $\pm$ 5.3 ab	72.0 $\pm$ 9.6 a	128.2 $\pm$ 21.1 a	100 $\pm$ 0
Both	34	42.9 $\pm$ 5.3 a	76.4 $\pm$ 12.2 a	126.6 $\pm$ 18.9 a	100 $\pm$ 0
<b>All lures</b>					
Light Green	68	26.6 $\pm$ 2.8 b	59.7 $\pm$ 7.5 a	95.8 $\pm$ 13.1 a	97 $\pm$ 2
Dark Purple	68	53.7 $\pm$ 4.6 a	51.6 $\pm$ 5.6 a	111.3 $\pm$ 11.4 a	100 $\pm$ 0
<i>F</i> ; <i>df</i> ; <i>P</i>					
Color		3.81; 1, 22; 0.05	0.49; 1, 22; 0.4	3.41; 1, 22; 0.08	
Lure		3.03; 3, 10; 0.03	7.67; 3, 10; 0.005	4.26; 3, 10; 0.03	
Color $\times$ lure (site)		1.71; 3, 10; 0.07	1.12; 3, 10; 0.4	1.2; 3, 10; 0.3	
Infestation level of tree		17.57; 1, 68; <0.0001	5.83; 1, 68; 0.02	9.64; 1, 68; 0.003	

Traps were hung in the canopies of ash trees along the edge of forest stands at three sites in Southern Michigan with varying emerald ash borer infestation levels in 2010. Mean numbers captured within a column followed by different letters are significantly different, Tukey test,  $P < 0.05$ .

**TABLE 3** | Mean number ( $\pm$  SE) of *A. planipennis* captured and detection rate (percentage of traps that captured at least one beetle) in Experiment 2 that compared different colored double-decker (DD) traps along the edge of a forest stand at Legg Park, Ingham County Michigan with a very low emerald ash borer population level in 2014.

Trap color and treatment	Mean number of EAB captured $\pm$ SE			Detection rate (%)
	Females	Males	Total	
Dark Purple DD HexHex	0.9 $\pm$ 0.4 b	1.1 $\pm$ 0.4 bc	2.0 $\pm$ 0.8 b	70.0 $\pm$ 15
Dark Purple DD HexMan	0.8 $\pm$ 0.3 b	1.0 $\pm$ 0.5 c	1.8 $\pm$ 0.8 b	60.0 $\pm$ 16
Light Purple DD HexHex	1.9 $\pm$ 0.5 ab	3.8 $\pm$ 1.5 ab	5.6 $\pm$ 1.9 a	90.0 $\pm$ 10
Green/Purple DD HexHex	3.1 $\pm$ 0.7 a	5.9 $\pm$ 0.9 a	8.9 $\pm$ 1.5 a	100.0 $\pm$ 0
Green/Purple DD HexMan	1.7 $\pm$ 0.5 ab	4.7 $\pm$ 1.1 a	6.4 $\pm$ 1.2 a	100.0 $\pm$ 0
<i>F</i> ; <i>df</i> ; <i>P</i>	4.17; 4, 36; 0.007	9.25; 4, 36; <0.0001	10.47; 4, 36; <0.0001	

Double-decker traps consisted of a 3-m PVC pole with an upper prism trap mounted to the top and lower prism trap mounted 60 cm below. Double-decker traps included dark purple for both the upper and lower prisms (Dark Purple), light purple for both the upper and lower prisms (Light Purple), or green for the upper prism and light purple for the lower prism (Green/Purple). Traps were baited with *cis*-3-hexenol lures (7.4 mg/day) on both panels (HexHex) or *cis*-3-hexenol on the upper panel and Manuka oil (50 mg/day) on the lower panel (HexMan). Mean numbers captured within a column followed by different letters are significantly different, Tukey test,  $P < 0.05$ .  $N = 10$ .

dark green sticky prism traps. Dark purple double-decker traps captured an intermediate number of females. Differences among treatments followed a similar pattern but were not significant for males. The detection rate was 100% for all traps except for the dark green sticky prism traps of which 80% captured at least one *A. planipennis* (Table 4).

The results for experiment 4 in 2015 at Legg Park were similar to those for experiment 3 and *A. planipennis* populations were similarly very low after the peak infestation had declined. We captured 379 *A. planipennis* adults in the experiment. Differences among treatments were only significant for females and dark green prism traps hung in the canopy captured fewer females than dark green funnel or green/purple double-decker traps. The detection rates were 100% for double-decker traps, 87.5% for dark green funnel traps, and 75% for dark green prism traps (Table 5).

## DISCUSSION

Our results indicate that prism traps and multiple funnel traps hung in the canopy of ash trees and double-decker traps were all effective at capturing *A. planipennis* and a high percentage of traps detected *A. planipennis* even at sites with very low infestation levels. However, mean catch of males and females varied among trap types, colors, and lures tested. At sites with high population levels, differences among treatments were less pronounced. In experiment 1 at the site with lowest population density, females were more attracted to purple prism traps hung in the canopy and to Manuka oil and Phoebe oil lures than to green prism traps hung in the canopy or *cis*-3-hexenol lures; whereas, males were significantly more attracted to green sticky prism traps in the canopy and *cis*-3-hexenol lures than to purple sticky prism traps or Manuka

**TABLE 4** | Mean number ( $\pm$  SE) of *A. planipennis* captured and detection rate (percentage of traps that captured at least one beetle) in Experiment 3 that compared traps of different types and colors at a woodlot near Ithaca, Gratiot County, Michigan with a high infestation level and at Fox Memorial Park, Eaton County, Michigan with a very low infestation level in 2014.

Trap color and treatment	Mean number of EAB captured $\pm$ SE			Detection rate (%)
	Females	Males	Total	
<b>ITHACA</b>				
Green Funnel Hex	39.8 $\pm$ 15.1 a	41.4 $\pm$ 16.6 a	81.2 $\pm$ 31.3 a	100 $\pm$ 0
Green Sticky Prism Hex	36.4 $\pm$ 8.7 a	44.6 $\pm$ 17.6 a	81.0 $\pm$ 22.6 a	100 $\pm$ 0
Green/Purple DD HexHex	12.8 $\pm$ 1.4 ab	27.4 $\pm$ 6.5 ab	40.2 $\pm$ 7.1 ab	100 $\pm$ 0
Dark Purple DD HexMan	10.2 $\pm$ 3.3 b	7.6 $\pm$ 2.5 b	17.8 $\pm$ 5.4 b	100 $\pm$ 0
<i>F</i> ; <i>df</i> ; <i>P</i>	6.26; 3, 16; 0.005	56; 3, 16; 0.008	5.95; 3, 16; 0.006	
<b>Fox Memorial Park</b>				
Green Funnel Hex	8.4 $\pm$ 3.4 a	7.6 $\pm$ 2.9	16.0 $\pm$ 6.0 a	100 $\pm$ 0
Green Sticky Prism Hex	1.6 $\pm$ 1.1 b	2.2 $\pm$ 1.0	3.8 $\pm$ 2.2 b	80 $\pm$ 20
Green/Purple DD HexHex	5.3 $\pm$ 1.5 a	6.8 $\pm$ 3.1	12.2 $\pm$ 4.0 a	100 $\pm$ 0
Dark Purple DD HexMan	4.0 $\pm$ 1.4 ab	4.2 $\pm$ 1.9	8.2 $\pm$ 3.2 a	100 $\pm$ 0
<i>F</i> ; <i>df</i> ; <i>P</i>	4.78; 3, 12; 0.02	1.76; 3, 12; 0.2	3.48; 3, 12; 0.04	

Trap types included green funnel traps (Green Funnel) or green sticky prism traps (Green Sticky Prism) hung in the canopies of ash trees and baited with *cis*-3-hexenol lures (7.4 mg/day) (Hex), and double-decker traps with green upper and light purple lower panels (Green/Purple DD) baited with *cis*-3-hexenol lures on both panels (HexHex) or dark purple upper and lower panels (Dark Purple DD) baited with *cis*-3-hexenol on the upper panel and manuka oil (50 mg/day) on the lower panel (HexMan). Mean numbers captured within a column followed by different letters are significantly different, Tukey test,  $P < 0.05$ .  $N = 5$  blocks per site.

**TABLE 5** | Mean number ( $\pm$  SE) of *A. planipennis* captured and detection rate (percentage of traps that captured at least one beetle) in Experiment 4 that compared traps of different types and colors at a Legg Park, Ingham County, Michigan with a very low emerald ash borer infestation level in 2015.

Trap color and treatment	Mean number of EAB captured $\pm$ SE			Detection rate (%)
	Females	Males	Total	
Green Funnel Hex	4.8 $\pm$ 1.9 a	7.8 $\pm$ 3.0	12.5 $\pm$ 4.6	87.5 $\pm$ 12.5
Green Sticky Prism Hex	1.3 $\pm$ 0.6 b	5.7 $\pm$ 2.4	6.9 $\pm$ 2.3	75.0 $\pm$ 16.4
Green/Purple DD HexHex	4.5 $\pm$ 1.2 a	9.5 $\pm$ 2.8	14.0 $\pm$ 3.8	100.0 $\pm$ 0
<i>F</i> ; <i>df</i> ; <i>P</i>	6.64; 2, 14; 0.009	2.52; 2, 14; 0.1	1.66; 2, 14; 0.2	

Trap types included green funnel traps (Green Funnel) or green sticky prism traps (Green Sticky Prism) hung in the canopies of ash trees and baited with *cis*-3-hexenol lures (7.4 mg/day) (Hex), and double-decker traps with green upper and light purple lower panels (Green/Purple DD) baited with *cis*-3-hexenol lures on both panels (HexHex). Means within a column followed by different letters are significantly different, Tukey test,  $P < 0.05$ ,  $N = 8$ .

and Phoebe oil lures. Previous studies have found that females were more attracted than males to purple prism traps hung in the canopy (Crook et al., 2009; Francese et al., 2010b, 2013a) and males were significantly more attracted than females to green prism traps baited with *cis*-3-hexenol (Grant et al., 2011; Crook et al., 2012; Ryall et al., 2012). The wavelength of purple is similar to the spectrum of tree bark, suggesting that females that typically spend time resting and ovipositing on the boles of ash trees (Rodríguez-Saona et al., 2007; Lelito et al., 2008) may be attempting to locate suitable host sites for oviposition. On the other hand, males typically hover around the canopies of ash trees and land and rest on leaves or females feeding in the canopy (Lelito et al., 2007, 2008; Rodríguez-Saona et al., 2007). Therefore, males may be more attracted to colors and volatiles associated with leaves in the canopy. Because more males were captured than females, overall more *A. planipennis* of both sexes were captured in light green than in dark purple sticky prism traps at the site with the lowest

infestation level. Similarly, Francese et al. (2010b) found more *A. planipennis* were captured in green than in dark purple prism traps hung in the mid-canopy of ash trees. Our finding that significantly more *A. planipennis* were captured in light purple double-decker traps than in dark purple double-decker traps also support previous studies that found light Sabic purple was more attractive to *A. planipennis* than dark purple (Francese et al., 2010a, 2013a).

There was no significant difference in numbers of *A. planipennis* males or females captured on double-decker traps baited with *cis*-3-hexenol on both prisms compared to traps baited with *cis*-3-hexenol on the upper prism and Manuka oil on the lower prism among double-decker traps of the same color. *cis*-3-Hexenol is readily available from chemical suppliers as a synthetic green leaf alcohol and is relatively inexpensive and consistent in quality and supply. On the other hand, Manuka oil and Phoebe oil are natural steam distillates that can vary from batch to batch and supply may be inconsistent

and limited. Although Phoebe oil has been shown to enhance attraction of *A. planipennis* to Manuka oil due to the presence of an additional antennally-active bark volatile (Crook et al., 2012), it was not available for inclusion in experiments 3 and 4 conducted in 2014 and 2015. Due to inconsistent and limited availability of natural oils, *cis*-3-hexenol would be preferred for operational use.

Differences in responses by males and females to different colors and lures under different conditions suggest that there is no single best trap color and lure. Double-decker traps incorporate both green and purple prisms at different heights which may attract beetles of both sexes. In all 3 experiments that included green/purple double-decker traps, they consistently had 100% detection rate.

The infestation level of trees in which sticky prism traps were hung was a significant covariate in the model. Not surprisingly, traps hung in trees ranked as heavily infested captured more beetles than traps hung in lightly infested trees. Similarly, Poland and McCullough (2014) found a significant correlation between number of *A. planipennis* captured and infestation level of trees in which traps were hung and infestation level of trees in the vicinity of traps. In operational detection programs where *A. planipennis* is not known to be established there are unlikely to be any heavily infested trees.

Results for experiments conducted at sites with low infestation levels are most relevant to what is needed for operational detection surveys. Fox Memorial Park in experiment 3, and Legg Park in experiment 4 had very low *A. planipennis* infestations after peak outbreaks had declined. Most mature ash trees had died; however, there were abundant ash saplings at the sites. Although environmental and host conditions were different from a newly infested site, the very low *A. planipennis* populations would be similar to a new infestation. At these sites, the green/purple double-decker traps and green funnel traps captured significantly more females than green sticky prism traps and had higher detection rates. Thus, they may be more effective for early detection programs.

Each of the trap designs has advantages and disadvantages. Sticky prism traps hung in the canopy are light, and easy to assemble and install. However, they are coated with glue, messy to handle, and cannot be reused. They are hung in the canopy and, therefore, require suitable host trees and extra equipment for hanging. Rope or string for hanging traps may become tangled, and the glue-coated prisms may become stuck in foliage making it difficult to raise and lower traps for checking. Double-decker traps are bulkier and although the T-posts and PVC pipes can be reused, the glue-coated prisms are messy and cannot be used again. In addition, they require twice as many prisms and lures as sticky prism traps hung in the canopy which increases per trap costs. They can be easily set up anywhere in the open near the edge of a stand and do not require suitable hosts, rope, or hanging equipment; however, they require custom assembly. Multiple funnel traps are not sticky and can be reused but are more expensive and must be hung in the canopy and, therefore, have some of the same challenges as sticky prism traps with hanging equipment and ropes that may become tangled. Collection cups

require the use of a liquid or insecticidal killing agent and must be collected from every 3 to 4 weeks or captured insects may decay.

All trap contents are collected from the cups of multiple funnel traps and are later sorted and identified by experts in the laboratory; whereas field crews must recognize and collect insects suspected to be *A. planipennis* from the trapping glue on sticky prism traps. Although insects retrieved from sticky prism traps are verified by experts in the laboratory, it may be more likely that a positive trap capture on sticky prism traps might be missed by field crews than in multiple funnel traps.

It is generally assumed that an increased number of beetles captured corresponds to a higher probability of detection. However, in our experiments that was not necessarily true. In experiment 3, at the heavily infested site near Ithaca, MI, green funnel and green prism traps hung in the canopy captured significantly more *A. planipennis* on average than double-decker traps. However, variability in trap catches for traps hung in the canopy was much greater than for double-decker traps, possibly reflecting variable infestation levels of trees in which they were hung. Numbers of *A. planipennis* captured ranged from 8 to 180 (95% difference) in dark green funnel traps and from 12 to 159 (92% difference) in green prism traps, compared to a range from 15 to 31 (51% difference) in green/purple double-decker traps. Overall, green/purple double-decker traps appeared to be most consistent and had 100% detection rates at sites with very low to high infestations in all 3 experiments in which they were tested. Other studies have also found that double-decker traps consistently have higher detection rates than other traps (Marshall et al., 2010a,b; McCullough et al., 2011; Poland and McCullough, 2014). Therefore, double-decker traps may be the most effective for detection and should be considered for areas where operational detection efforts are focused.

Many factors may interact and influence detection rate and number of *A. planipennis* captured: trap type, color, lure, trap placement within the canopy or in the open along the forest edge, sun exposure, ash host tree density, stand condition, infestation level of the tree in which the trap is hung, and *A. planipennis* population density. These and other factors should be considered in selection of trap type for detection surveys. Future research is needed to determine the effective trapping radius of different trap types and their accuracy in reflecting infestation level of a stand and attack densities in surrounding trees. Positive detection of *A. planipennis* in a trap currently does not provide detailed information about proximity of infested trees or site infestation levels, although number captured may provide an indication of general infestation level. Operational programs must also consider logistical issues including trap costs and longevity. While sticky prisms are initially less expensive than multiple funnel traps, they must be replaced each year, while multiple funnel traps may be reused for several decades. Future research is also needed to determine the cost benefit of different trap types and the optimal density of traps to deploy over a given survey area in relation to host density.

## DATA AVAILABILITY STATEMENT

The datasets generated for this study are available on request to the corresponding author.

## AUTHOR CONTRIBUTIONS

TPo analyzed the data and wrote the first draft of the manuscript. TPo, TPe, and TC helped design the studies, conduct the field experiments, and edited the manuscript.

## REFERENCES

- Allison, J. D., Graham, E. E., Poland, T. M., and Strom, B. L. (2016). Dilution of fluon before trap surface treatment has no effect on longhorned beetle (Coleoptera: Cerambycidae) captures. *J. Econ. Entomol.* 109, 1215–1219. doi: 10.1093/jee/tow081
- Bartels, D., Williams, D., Ellenwood, J., and Sapio, F. (2008). “Accuracy assessment of remote sensing imagery for mapping hardwood trees and emerald ash borer-stressed ash trees,” in *Emerald Ash Borer Research and Technology Development Meeting, 23–24 October 2007, Pittsburgh, PA: Department of Agriculture, Forest Service Publication FHTET-2008-07*, eds V. Mastro, D. Lance, R. Reardon, and G. Parra (Morgantown, WV), 63–65.
- Brockerhoff, E. G., Liebhold, A. M., Richardson, B., and Suckling, D. M. (2010). Eradication of invasive forest insects: concepts, methods, costs and benefits. *N. Z. J. For. Sci.* 40(Suppl.), S117–S135.
- Burr, S. J., McCullough, D. G., and Poland, T. M. (2018). Density of emerald ash borer (Coleoptera: Buprestidae) adults and larvae at three stages of the invasion wave. *Environ. Entomol.* 47, 121–132. doi: 10.1093/ee/nvx200
- CFIA (2019). *Emerald Ash Borer – Agrilus Planipennis*. Available Online at: <https://www.inspection.gc.ca/plants/plant-pests-invasive-species/insects/emerald-ash-borer/eng/1337273882117/1337273975030>
- Crook, D., Khrimian, A., Francese, J. A., Fraser, I., Poland, T. M., Sawyer, A. J., et al. (2008). Development of a host-based semiochemical lure for trapping emerald ash borer, *Agrilus planipennis* (Coleoptera: Buprestidae). *Environ. Entomol.* 37, 356–365. doi: 10.1093/ee/37.2.356
- Crook, D. J., Francese, J. A., Rietz, M. L., Lance, D. R., Hull-Sanders, H. M., Mastro, V. C., et al. (2014). Improving detection tools for emerald ash borer (Coleoptera: Buprestidae): comparison of multifunnel traps, prism traps, and lure types at varying population densities. *J. Econ. Entomol.* 107, 1496–1501. doi: 10.1603/EC14041
- Crook, D. J., Francese, J. A., Zylstra, K. E., Fraser, I., Sawyer, A. J., Bartels, D. W., et al. (2009). Laboratory and field response of the emerald ash borer (Coleoptera: Buprestidae) to selected regions of the electromagnetic spectrum. *J. Econ. Entomol.* 102, 2160–2169. doi: 10.1603/029.102.0620
- Crook, D. J., Khrimian, A., Cossé, A., Fraser, I., and Mastro, V. C. (2012). Influence of trap color and host volatiles on capture of the emerald ash borer (Coleoptera: Buprestidae). *J. Econ. Entomol.* 105, 429–437. doi: 10.1603/EC11204
- Crook, D. J., and Mastro, V. C. (2010). Chemical ecology of the emerald ash borer *Agrilus planipennis*. *J. Chem. Ecol.* 36, 101–112. doi: 10.1007/s10886-009-9738-x
- de Groot, P., Grant, G. G., Poland, T. M., Scharbach, R., Buchan, L., Nott, R. W., et al. (2008). Electrophysiological response and attraction of emerald ash borer to green leaf volatiles (GLVs) emitted by host foliage. *J. Chem. Ecol.* 34, 1170–1179. doi: 10.1007/s10886-008-9514-3
- Domingue, M. J., Lelito, J. P., Fraser, I., Mastro, V. C., Tumlinson, J. H., and Baker, T. C. (2013). Visual and chemical cues affecting the detection rate of the emerald ash borer in sticky traps. *J. Appl. Entomol.* 137, 77–87. doi: 10.1111/j.1439-0418.2012.01737.x
- Domingue, M. J., Pulsifer, D. P., Lakhtakia, A., Berkebile, J., Steiner, K. C., Lelito, J. P., et al. (2015). Detecting emerald ash borers (*Agrilus planipennis*) using branch traps baited with 3D-printed beetle decoys. *J. Pest Sci.* 88, 267–279. doi: 10.1007/s10340-014-0598-y

## FUNDING

This research was supported by federal funds from the USDA Forest Service.

## ACKNOWLEDGMENTS

We thank Tom Baweja, Emily Loubert, and Elizabeth Graham for assistance with field experiments, John Stanovick for assistance with statistical analyses, and Robert Haack, Dan Miller, and two reviewers for comments on an earlier version of the manuscript.

- Fierke, M. K., Whitmore, M. C., Foelker, C., Vandenberg, J. D., and Carlson, J. (2013). Delimitation and management of emerald ash borer (Coleoptera: Buprestidae): case study at an outlier infestation in southwestern New York State, United States of America. *Can. Entomol.* 145, 577–587. doi: 10.4039/tce.2013.39
- Francese, J. A., Booth, E. G., Lopez, V. M., and Sorensen, B. (2019). Alternative survey methods for the emerald ash borer. *FL. Entomol.* 102, 243–245. doi: 10.1653/024.102.0142
- Francese, J. A., Crook, D. J., Fraser, I., Lance, D. R., Sawyer, A. J., and Mastro, V. C. (2010a). Optimization of trap color for the emerald ash borer, *Agrilus planipennis* (Coleoptera: Buprestidae). *J. Econ. Entomol.* 103, 1235–1241. doi: 10.1603/EC10088
- Francese, J. A., Fraser, I., Lance, D. R., and Mastro, V. C. (2011). Efficacy of multifunnel traps for capturing emerald ash borer (Coleoptera: Buprestidae): effect of color, glue, and other trap coatings. *J. Econ. Entomol.* 104, 901–908. doi: 10.1603/EC11038
- Francese, J. A., Fraser, I., Rietz, M. L., Crook, D. J., Lance, D. R., and Mastro, V. C. (2010b). Relation of color, size, and canopy placement of prism traps in determining capture of emerald ash borer (Coleoptera: Buprestidae). *Can. Entomol.* 142, 596–600. doi: 10.4039/n10-041
- Francese, J. A., Oliver, J. B., Fraser, I., Lance, D. R., Youssef, N., Sawyer, A. J., et al. (2008). Influence of trap placement and design on capture of the emerald ash borer (Coleoptera: Buprestidae). *J. Econ. Entomol.* 101, 1831–1837. doi: 10.1603/0022-0493-101.6.1831
- Francese, J. A., Rietz, M. L., Crook, D. J., Fraser, I., Lance, D. R., and Mastro, V. C. (2013a). Improving detection tools for the emerald ash borer (Coleoptera: Buprestidae): comparison of prism and multifunnel traps at varying population densities. *J. Econ. Entomol.* 106, 2407–2414. doi: 10.1603/EC13013
- Francese, J. A., Rietz, M. L., and Mastro, V. C. (2013b). Optimization of multifunnel traps for emerald ash borer (Coleoptera: Buprestidae): influence of size, trap coating and color. *J. Econ. Entomol.* 106, 2415–2423. doi: 10.1603/EC13014
- Graham, E. E., and Poland, T. M. (2012). Efficacy of fluon conditioning for capturing cerambycid beetles in different trap designs and persistence on panel traps over time. *J. Econ. Entomol.* 105, 395–401. doi: 10.1603/EC11432
- Grant, G. G., Poland, T. M., Ciaramitaro, T., Lyons, D. B., and Jones, G. C. (2011). Comparison of male and female emerald ash borer (Coleoptera: Buprestidae) responses to Phoebe oil and (z)-3-hexenol lures in light green prism traps. *J. Econ. Entomol.* 104, 173–179. doi: 10.1603/EC10197
- Grant, G. G., Ryall, K. L., Lyons, D. B., and Abou-Zaid, M. M. (2010). Differential response of male and female emerald ash borers (Col., Buprestidae) to (Z)-3-hexenol and manuka oil. *J. Appl. Entomol.* 134, 1–8. doi: 10.1111/j.1439-0418.2009.01441.x
- Haack, R. A., Baranchikov, Y., Bauer, L. S., and Poland, T. M. (2015). “Chapter 1: emerald ash borer biology and invasion history,” in *Biology and Control of Emerald Ash Borer*, eds R. G. Van Driesche and R. C. Reardon (Morgantown, WV: USDA Forest Service, Forest Health Technology Enterprise Team, FHTET-2014-09), 1–13.
- Haack, R. A., Jendek, E., Liu, H., Marchant, K. R., Petrice, T. R., Poland, T. M., et al. (2002). The emerald ash borer: a new exotic pest in North America. *Newsl. Mich. Entomol. Soc.* 47, 1–5. Available online at: [https://www.nrs.fs.fed.us/pubs/jrnl/2002/nc\\_2002\\_haack\\_001.pdf](https://www.nrs.fs.fed.us/pubs/jrnl/2002/nc_2002_haack_001.pdf)

- Herms, D. A., and McCullough, D. G. (2014). Emerald ash borer invasion of North America: history, biology, ecology, impacts, and management. *Ann. Rev. Entomol.* 59, 13–30. doi: 10.1146/annurev-ento-011613-162051
- Lelito, J. P., Böröczky, K., Jones, T. H., Fraser, I., Mastro, V. C., Tumlinson, J. H., et al. (2009). Behavioral evidence for a contact sex pheromone component of the emerald ash borer, *Agrilus planipennis* Fairmaire. *J. Chem. Ecol.* 35, 104–110. doi: 10.1007/s10886-008-9583-3
- Lelito, J. P., Fraser, I., Mastro, V. C., Tumlinson, J. H., and Baker, T. C. (2008). Novel visual-cue-based sticky traps for monitoring of emerald ash borers, *Agrilus planipennis* (Col., Buprestidae). *J. Appl. Entomol.* 132, 668–674. doi: 10.1111/j.1439-0418.2008.01308.x
- Lelito, J. P., Fraser, I., Mastro, V. C., Tumlinson, J. H., Böröczky, K., and Baker, T. C. (2007). Visually mediated 'paratrooper copulations' in the mating behavior of *Agrilus planipennis* (Coleoptera: Buprestidae), a highly destructive invasive pest of North American ash trees. *J. Insect Behav.* 20, 537–552. doi: 10.1007/s10905-007-9097-9
- Liebold, A. M., and Tobin, P. C. (2008). Population ecology of insect invasions and their management. *Ann. Rev. Entomol.* 53: 387–408. doi: 10.1146/annurev.ento.52.110405.091401
- Lyons, J. E., Runge, M. C., Laskowski, H. P., and Kendall, W. L. (2008). Monitoring in the context of structured decision-making and adaptive management. *J. Wildl. Manag.* 72, 1683–1692. doi: 10.2193/2008-141
- Marshall, J. M., Storer, A. J., Fraser, I., Beachy, J. A., and Mastro, V. C. (2009). Effectiveness of differing trap types for the detection of emerald ash borer (Coleoptera: Buprestidae). *Environ. Entomol.* 38, 1226–1234. doi: 10.1603/022.038.0433
- Marshall, J. M., Storer, A. J., Fraser, I., and Mastro, V. C. (2010a). Efficacy of trap and lure types for detection of *Agrilus planipennis* (Col., Buprestidae) at low density. *J. Appl. Entomol.* 134, 296–302. doi: 10.1111/j.1439-0418.2009.01455.x
- Marshall, J. M., Storer, A. J., Fraser, I., and Mastro, V. C. (2010b). "Multi-state comparison of detection tools at low emerald ash borer densities," in *Proceedings of the Emerald Ash Borer and Asian Longhorned Beetle Research and Technology Development Meeting, Pittsburgh, PA, 19-21 Oct. 2009*, eds V. Mastro, D. Lance, R. Reardon, and G. Parra (Morgantown, WV: U.S. Department of Agriculture, Forest Service Publication), 124–125.
- McCullough, D. G., Poland, T. M., Anulewicz, A. C., and Cappaert, D. (2009a). Emerald ash borer (*Agrilus planipennis* Fairmaire) (Coleoptera: Buprestidae) attraction to stressed or baited ash (*Fraxinus* spp.) trees. *Environ. Entomol.* 38, 1668–1679. doi: 10.1603/022.038.0620
- McCullough, D. G., Poland, T. M., Cappaert, D., and Anulewicz, A. C. (2009b). Emerald ash borer (*Agrilus planipennis*) attraction to ash trees stressed by girdling, herbicide and wounding. *Can. J. For. Res.* 39, 1331–1345. doi: 10.1139/X09-057
- McCullough, D. G., Siegert, N. W., Poland, T. M., Pierce, S. J., and Ahn, S. Z. (2011). Effects of trap type, placement and ash distribution on emerald ash borer captures in a low density site. *Environ. Entomol.* 40, 1239–1252. doi: 10.1603/EN11099
- Musolin, D. L., Selikhovkin, A. V., Shabunin, D. A., Zviagintsev, V. B., and Baranchikov, Y. N. (2017). Between ash dieback and emerald ash borer: two Asian invaders in Russia and the future of ash in Europe. *Baltic For.* 23, 316–333.
- Orlova-Bienkowskaja, M. J., Drogvalenko, A. N., Zabaluev, I. A., Sazhnev, A. S., Peregudova, H. Y., Mazurov, S. G., et al. (2019). Bad and good news for ash trees in Europe: alien pest *Agrilus planipennis* has spread to the Ukraine and the south of European Russia, but does not kill *Fraxinus excelsior* in the forests. *BioRxiv*. doi: 10.1101/689240
- Orlova-Bienkowskaja, M. J. (2014). Ashes in Europe are in danger: the invasive range of *Agrilus planipennis* in European Russia is expanding. *Biol. Inv.* 16, 1345–1349. doi: 10.1007/s10530-013-0579-8
- Petrice, T. R., Haack, R. A., and Poland, T. M. (2013). Attraction of *Agrilus planipennis* (Coleoptera: Buprestidae) and other buprestids to sticky traps of various colors and shapes. *Gt. Lakes Entomol.* 46, 13–30. Available online at: [https://www.fs.fed.us/nrs/pubs/jrnl/2013/nrs\\_2013\\_petrice\\_001.pdf](https://www.fs.fed.us/nrs/pubs/jrnl/2013/nrs_2013_petrice_001.pdf)
- Poland, T. M., and McCullough, D. G. (2006). Emerald ash borer: Invasion of the urban forest and the threat to North America's ash resource. *J. For.* 104, 118–124. doi: 10.1093/jof/104.3.118
- Poland, T. M., and McCullough, D. G. (2014). Comparison of trap types and colors for capturing emerald ash borer adults at different population densities. *Environ. Entomol.* 43, 157–170. doi: 10.1603/EN13137
- Poland, T. M., McCullough, D. G., and Anulewicz, A. C. (2011). Evaluation of double-decker traps for emerald ash borer (Coleoptera: Buprestidae). *J. Econ. Ent.* 104, 517–531. doi: 10.1603/EC10254
- Poland, T. M., and Rassati, D. (2018). Improved early detection of invasive forest insects – a review of current methods and research. *J. Pest. Sci.* 92, 37–49. doi: 10.1007/s10340-018-1004-y
- Rassati, D., Faccoli, M., Petrucco Toffolo, E., Battisti, A., and Marini, L. (2015). Improving the early detection of alien wood-boring beetles in ports and surrounding forests. *J. Appl. Ecol.* 52, 50–58. doi: 10.1111/1365-2664.12347
- Rodriguez-Saona, C., Poland, T. M., Miller, J. R., Stelinski, L. L., Grant, G. G., de Groot, P., et al. (2006). Behavioral and electrophysiological responses of the emerald ash borer, *Agrilus planipennis*, to induced volatiles of Manchurian ash, *Fraxinus mandshurica*. *Chemoecol.* 16, 75–86. doi: 10.1007/s00049-005-0329-1
- Rodriguez-Saona, C. R., Miller, J. R., Poland, T. M., Kuhn, T. M., Otis, G. W., Turk, T., et al. (2007). Behaviours of adult emerald ash borer, *Agrilus planipennis* (Coleoptera: Buprestidae). *Gt. Lakes Entomol.* 40, 1–16. Available online at: <https://scholar.valpo.edu/tgle/vol40/iss1/>
- Ryall, K. (2015). Detection and sampling of emerald ash borer (Coleoptera: Buprestidae) infestations. *Can. Entomol.* 147, 290–299. doi: 10.4039/tce.2015.7
- Ryall, K. L., Fidge, J. G., Silk, P. J., and Scarr, T. A. (2013). Efficacy of the pheromone (3Z)-lactone and the host kairomone (3Z)-hexenol at detecting early infestation of the emerald ash borer, *Agrilus planipennis*. *Entomol. Exp. Appl.* 147, 126–131. doi: 10.1111/eea.12052
- Ryall, K. L., Silk, P. J., Mayo, P., Crook, D., Khirman, A., and Cossé, A. A. (2012). Attraction of *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae) to a volatile pheromone: effects of release rate, host volatile and trap placement. *Environ. Entomol.* 41, 648–656. doi: 10.1603/EN11312
- SAS Institute (2012). *OC User's Manual, Version 9.4*. Cary, NC: SAS Institute.
- Siegert, N. W., McCullough, D. G., Liebold, A. M., and Telewski, F. W. (2014). Dendrochronological reconstruction of the epicentre and early spread of emerald ash borer in North America. *Div. Distrib.* 20, 847–858. doi: 10.1111/ddi.12212
- Silk, P. J., Ryall, K., Lyons, D. B., Sweeney, J., and Wu, J. (2009). A contact sex pheromone component of the emerald ash borer, *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae). *Naturwiss* 96, 601–608. doi: 10.1007/s00114-009-0513-1
- Silk, P. J., Ryall, K., Mayo, P., Lemay, M., Grant, G. G., and Crook, D. (2011). Evidence for a volatile pheromone in *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae) that increases attraction to a host foliar volatile. *Environ. Entomol.* 40, 904–916. doi: 10.1603/EN11029
- Silk, P. J., Ryall, K., and Roscoe, L. (2019). Emerald ash borer, *Agrilus planipennis* (Coleoptera: Buprestidae), detection and monitoring in Canada. *For. Int. J. For. Res.* 10, 323–343. doi: 10.1093/forestry/cpz036
- Tobin, P. C., Kean, J. M., Suckling, D. M., McCullough, D. G., Herms, D. A., and Stringer, L. D. (2014). Determinants of successful arthropod eradication programs. *Biol. Invasions.* 16, 401–414. doi: 10.1007/s10530-013-0529-5
- USDA-APHIS-PPQ (2018). *2018 Emerald Ash Borer Survey Guidelines*. Available online at: [http://www.aphis.usda.gov/plant\\_health/plant\\_pest\\_info/emerald\\_ash\\_b/downloads/survey\\_guidelines.pdf](http://www.aphis.usda.gov/plant_health/plant_pest_info/emerald_ash_b/downloads/survey_guidelines.pdf)
- Yu, C. (1992). "Agrilus marcopoli Obenberger," in *Forest Insects of China, 2nd Edn*, ed G. Xiao (Beijing: China Forestry Publishing House), 400–401.

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Poland, Petrice and Ciaramitaro. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.