

Impact of Exotic Earthworms on Organic Carbon Sorption on Mineral Surfaces and Soil Carbon Inventories in a Northern Hardwood Forest

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ABSTRACT

Exotic earthworms are invading forests in North America where native earthworms have been absent since the last glaciation. These earthworms bioturbate soils and may enhance physical interactions between minerals and organic matter (OM), thus affecting mineral sorption of carbon (C) which may affect C cycling. We quantitatively show how OM-mineral sorption and soil C inventories respond to exotic earthworms along an earthworm invasion chronosequence in a sugar maple forest in northern Minnesota. We hypothesized that mineral surface area in A horizons would increase as burrowing earthworms incorporated clay minerals from the B

horizons and that enhanced contacts between OM and minerals would increase the OM sorption on mineral surfaces and mineral-associated C inventories in A horizons. Contrary to our hypotheses, mineral surface areas within A horizons were lowered because earthworm burrows only extended into the silt-rich loess that separated the A and clay-rich B horizons. Furthermore, where endogeic earthworms were present, a smaller fraction of mineral surface area was covered with OM. OM sorption on minerals in the A horizons shifted from a limitation of mineral surface availability to a limitation of OM availability within a decade after the arrival of endogeic earthworms. C-mineral sorption depends on earthworm consumption of OM as well as the composition and vertical distribution of minerals. This finding may thus explain the contradictory results reported in earlier investigations. Our results highlight the rapid and drastic effects of exotic earthworms on key ecosystem processes in deciduous forests in post-glacial settings.

Key words: biological invasion; earthworms; bioturbation; soil carbon; organic matter; minerals; exotic species; sorption; mineral surface area.

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Author contributions A. Lyttle conducted part of laboratory analysis, analyzed data, and wrote the first draft. K. Yoo conceived and designed the study, led collaborative field and laboratory research, analyzed data, and wrote the paper. C. Hale supervised earthworm collection and identification. A. Aufdenkampe contributed to designing BET surface area analysis, study design, and sample collection. S. D. Sebestyen contributed to study design and writing. K. Resner contributed to laboratory analysis. A. Blum conducted quantitative mineralogical analysis.

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INTRODUCTION

Earthworms are common in temperate and tropical ecosystems (Hendrix and others 2008) and among the best-known ecosystem engineers (Crooks 2002). Although they are not native species, exotic earthworms have invaded previously glaciated forests in North America (Hendrix and others 2008). Over past centuries, human activities such as agriculture, dumping of unused fishing bait, logging, recreation, transport of earthworm, and cocoon-infested soils have dramatically accelerated the areal expansion of exotic earthworms into post-glacial soils (for example, Hendrix and Bohlen 2002). Once introduced, exotic earthworms have substantially modified soil morphology and nutrient cycling (Alban and Berry 1994; Scheu and Parkinson 1994a, b; Bohlen and others 2004a, c; Frelich and others 2006; Hale and others 2008). In northern hardwood forests in North America, exotic earthworms have had effects on soil carbon (C) and nitrogen cycling (Hale and others 2005a; Frelich and others 2006; Bohlen and others 2004b, c; Hale and others 2008) and caused the decline of native understory plant communities (Alban and Berry 1994; Scheu and Parkinson 1994b; Gundale 2002; Bohlen and others 2004c; Hale and others 2005a, b, 2006).

Studies generally find that exotic earthworms cause a loss of the leaf litter layer in post-invasion forests. Nonetheless there are contrasting effects of exotic earthworms on C inventories within mineral soils that may arise from differences among locations and stages of earthworm invasion, highlighting a need for a process-based understanding of the effects of exotic earthworms on soil C turnover. Decades of research have converged on a consensus that long-term retention of OM in soils is primarily controlled by OM-mineral interactions (for example, Torn and others 1997; Schmidt and others 2011). Minerals protect C by occluding OM and together they cohere to form mineral aggregates (Baldock and Skjemstad 2000; von Lutzow and others 2006).

Here we focus on OM sorption on minerals surface as a key ecosystem process that may be strongly affected by exotic earthworms. To sorb organic matter onto mineral surfaces, minerals and organic matter must have physical contact, which requires turbation. Nonetheless, little is known about how this process is altered by bioturbation. Plant C inputs and C concentrations are typically high in surface soils, whereas secondary clay minerals presumably with high specific surface area (SSA) tend to be more abundant in illuvial

horizons at deeper depths. Therefore we conceptualize that many soils have a capacity for OM-mineral sorption that is limited by physical contact and that bioturbation may mix OM in the forest floor and clay minerals in subsoils, facilitating OM-mineral sorption. This basic premise underlies our approach to scale up nano-scale C-mineral sorption (for example, Remusat and others 2012) to whole-ecosystem processes that determine soil C cycling and budgets.

We studied an earthworm invasion chronosequence in a sugar maple forest of northern Minnesota. Along this chronosequence, different functional groups of earthworms have longitudinally progressed at different rates, and systematic responses of soil C contents and N and P availability to arrivals of different earthworm functional groups have been well established (Hale and others 2005a, b, Hale and others 2006). In exploring effects of earthworm bioturbation on C-mineral sorption, this detailed background information allowed us to systematically develop a soil sampling scheme that would balance:

- Existing knowledge of the distribution of earthworm functional groups and invasion sequence,
- Labor-intensive deep soil sampling, which severely limited sample size, and,
- Multi-faceted laboratory analyses, which were labor and time consuming.

We attempted to better understand the mechanisms and changes in magnitudes of OM sorption to mineral surface along an earthworm invasion chronosequence. We tested two specific hypotheses: (1) Burrowing earthworms increase mineral SSA in the A horizon by incorporating clay minerals from the underlying clay-rich B horizons; (2) earthworms that actively mix the A horizon increase OM sorption on mineral surfaces and thus increase mineral-sorbed soil C inventories.

METHODS

Study Site

The study site was a hardwood forest dominated by sugar maple (*Acer saccharum*) and basswood (*Tilia americana*) within the Chippewa National Forest in Northern Minnesota, USA. The climate is humid, continental, and cold temperate, with a mean annual temperature of 4 °C (Minnesota State Climatology Office 2003). The mean annual precipitation is 50–65 cm, with the growing season varying between 100 and 120 days (Adams and others 2004). The surficial geology of the site includes Guthrie

glacial till overlain by a silt loam loess layer of varying depths (Adams and others 2004). The land surface is gently rolling and seasonal wetlands are present in some depressions.

Earthworm-free soils had an O horizon and a thin A horizon. The forest litter layer was over 5 cm thick in earthworm-free areas. The A horizons, rarely exceeding 5 cm in thickness in earthworm-free areas, had many fine roots. Prior to earthworm invasion, the A horizon materials had a weak very fine granular structure and very fine sandy loam texture. The A horizon was underlain by a thick (~30 cm) silty layer with uniform thickness, mineralogy, texture, and pH which we refer to as loess. Between the loess layer and the underlying calcareous glacial till, we found a clay-rich B horizon. The loess materials had a very fine sandy loam texture and fine roots were present. Depths to carbonate rich glacial till ranged from 90 to 160 cm. The 200-m long transect extended from an earthworm-infested area located 145 m away from a secondary road into the forest where earthworms were nearly absent (Figure 1).

Soil Sampling

Large soil pits (1.5 × 1.5 × 1.5 m) were excavated in 2009 to 20–30 cm below soil-glacial till boundaries (~100 to 150-cm depths) at 0, 50, 100, 150, 160, and 190 m along a transect (Figure 1). Soil samples were collected from the excavated pits in 2.5-cm increments in the A horizon, 5-cm increments in the loess layer, and 10-cm increments in

the B horizon. Bulk densities were determined for core samples from each horizon that were collected using a sliding hammer corer, dried, and weighed. In 2011, soil cores with internal diameters of 3.2 cm and lengths of 30 cm were collected adjacent to all plots along transect B (Figure 1) to measure thicknesses of A horizons and loess layers.

Earthworm Sampling and Identification

Earthworm biomass and speciation were quantified at 10-m intervals along transect B (Figure 1) in 2009. Earthworms were collected from 0.3-m² subplots using liquid mustard extraction (Lawrence and Bowers 2002; Hale and others 2005a). We sampled earthworms during September when earthworms were sexually mature and easier to identify. Collected earthworms were euthanized in 70 % isopropyl and preserved in 10 % formalin. Earthworm taxonomy was determined following Schwert (1990), Reynolds (1977), and Hale (2007). Ash-free dry (AFD) earthworm biomasses were calculated from the regression model developed by Hale and others (2004).

Earthworms were classified into three functional groups based upon feeding and dwelling habits. The epigeic group fed and dwelled in leaf litter without burrowing into mineral soils. Earthworms that dwelled within shallow mineral soils were in the endogeic functional group. These earthworms ingested mineral soil and digested organic matter. The endogeic group also created a system of connected non-permanent burrows in the mineral soil.

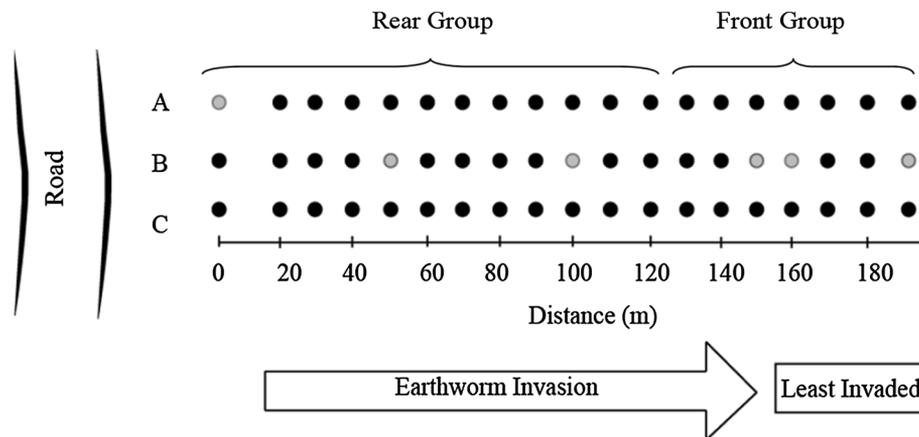


Figure 1. Earthworm invasion chronosequence. *Grey circles* show the soil pits that were excavated in 2009. Of all the locations shown with *black circles* and studied by Hale and others (2005b), 19 points along transect B were used for earthworm sampling in 2009 and were probed in 2011 to measure A horizon thicknesses. The soil pit at 0 meter was excavated in the transect A to avoid a wetland. The start of the transect (that is, Distance = 0 m) was 145 m from the road. The figure includes the road only to illustrate the source area and direction of exotic earthworms.

Finally, anecic earthworms fed on surface litter and pulled litter into their deep permanent burrows in mineral soils. Various earthworm species and their biomasses within each functional group at this site were described in Hale and others (2005b).

Carbon Content Measurements and Density Fractionation

To quantify mineral-associated C inventories—in addition to total C inventories—in A horizons, eighteen soil samples (~3 samples within the A horizons at each pit) across the invasion gradient were selected for density fractionation. Samples were wet sieved (2 mm), which may disintegrate macroaggregates. This process, however, would not interfere with our focus on mineral sorption of C instead of C occlusion in aggregates. Regardless, the coarse fraction (>2 mm) was less than 5 % (by mass) of samples. Approximately 10 g of the fine fraction (<2 mm) samples were air-dried, lightly ground using a rolling pin for 3 min, and dry sieved (250 μm) to selectively remove fine roots. Soil samples were then mixed with 2.0 g cm^{-3} sodium polytungstate (Torresan 1987) and placed on a shaker for 2–3 h at room temperature. Based on Sollins and others (2009), a density of 2.0 g cm^{-3} was used to remove organic debris and to focus on OM associated with mineral surfaces. Settled materials were retrieved and rinsed with deionized water until electrical conductivity of the solution was $\leq 50 \mu\text{S cm}^{-1}$ (Eusterhues and others 2005). The samples were then oven dried at 60 °C.

Oven-dried soil samples were treated with hydrochloric acid to remove carbonate prior to analysis. Soil organic C contents for bulk and density fractionated soil samples were determined by a VarioMax CN Elemental Analyzer at the Agricultural Research Station (USDA-ARS) facility at the University of Minnesota. Carbon contents in soil and leaf litter were measured on duplicate 0.5 g samples.

Specific Surface Area Measurements

Specific mineral surface area (SSA) was measured for fine fraction (<2 mm) soil samples using a TriStar 3020 Surface Area and Porosity Analyzer in the Department of Soil, Water, and Climate at the University of Minnesota. Based on the Brunauer–Emmett–Teller (BET) theory (Webb and Orr 1997), isotherm data were collected at 11 different pressure points at the boiling temperature of liquid nitrogen (that is, 77 K), using N_2 as an adsorbent

gas. Each sample weighed approximately 1.5 g. Samples were degassed prior to analysis to remove any excess water and contaminants on the surfaces: samples were heated at 150 °C and purged by N_2 gas for 4–6 h.

After initial SSA analysis of untreated soil, OM was removed from the same samples to reveal the mineral surface covered by OM. The OM-removed samples were then subject to the same SSA analysis. For removing OM from soil or sediment samples, we adopted the procedures used in Wagai and others (2009) where OM is removed by muffling samples at 350 °C for 12 h.

Iron and Aluminum Oxides Extractions

Because pedogenic crystalline and amorphous Fe/Al oxides have large specific surface areas (Eusterhues and others 2005; Kaiser and Guggenberger 2007) and affect carbon turnover (Torn and others 1997), we measured extractable Fe and Al on selected soil samples from heavily invaded (0 and 50 m) and minimally invaded (160 and 190 m) soils. Samples from three depths in the A horizons were analyzed. Soil samples of 0.75 g were mixed with sodium dithionite, sodium citrate, and sodium bicarbonate to remove both crystalline and non-crystalline Fe/Al oxides in the soil (Holmgren 1967). Extracts were then analyzed with an Inductively Coupled Plasma Atomic Emission Spectrometer (ICP-AES) at the Research Analytical Laboratory at the University of Minnesota (<http://ral.cfans.umn.edu/>). The non-crystalline Fe oxide pool was estimated by ammonium oxalate dark extraction (McKeague and Day 1966). Soil samples of 0.5 g were mixed with 50 mL of ammonium oxalate at pH 3 in dark bottles and were placed on shaker for 4 h. Later, samples were centrifuged and aliquots of the extracts were analyzed by ICP-AES.

Quantitative XRD

To constrain the mineralogical source of surface area, 1-g soil samples were ground with a zinc oxide standard. Using a Siemens D500 instrument at the United States Geological Survey, Boulder, CO, XRD spectra were collected from 5° to 65° 2θ with Cu and $\text{K}\alpha$ radiation. Using the RockJock program, mineral concentrations were determined according to Eberl (2003). Given that errors associated with quantitative XRD are difficult to estimate, we interpret the results to be rough estimates.

CALCULATIONS

Mass Equivalent Depths (MED)

Because exotic earthworms affect soil bulk density (Hale and others 2005a), different masses of overlying soil materials may be found at identical soil depths. To eliminate this effect, we adopted mass equivalent depth (MED). The MED represents the overlying soil mass. For example, MED of 1.9 g cm^{-2} represents a soil depth where the overlying soil mass per cm^2 is 1.9 g which was equivalent to the thickness of pre-existing A horizons ($\sim 6 \text{ cm}$) prior to the arrival of epi-endogeic or endogeic species. An MED of 18.2 g cm^{-2} corresponded to the lower boundaries of the loess layer ($\sim 35\text{-cm}$ depths) and thus included A horizons thickened by endogeic mixing. The MED, in units of g cm^{-2} , was calculated as:

$$\text{MED} = \sum_{i=1}^N (\Delta z_i \times \text{BD}_i) \quad (1)$$

where BD was the bulk density (g cm^{-3}), Δz was the thickness of a sampled layer (cm), i indicated the i -th layer sampled, and N was the total number of sampled layers above the depth of interest.

Carbon Inventories

Soil C inventory for a horizon, X, was calculated as:

$$\text{CI}_X = \sum_{i=1}^N (c_i \times \text{MED}_i) \quad (2)$$

where CI_X was the C inventory of a soil horizon, X [kg C m^{-2}], and c was the mass fraction of C in a sample [kg kg^{-1}]. Coarse contents were negligible because the values were less than 5 % in all samples.

Specific Surface Areas

Mineral specific surface area (SSA) can be subdivided as:

$$\text{SSA}_{\text{total}} = \text{SSA}_{\text{occluded}} + \text{SSA}_{\text{exposed}} \quad (3)$$

where $\text{SSA}_{\text{total}}$ equaled the total SSA of the mineral, $\text{SSA}_{\text{occluded}}$ was the area covered by OM, and $\text{SSA}_{\text{exposed}}$ represented the area not covered by OM. Operationally, $\text{SSA}_{\text{total}}$ corresponds to the mineral SSA determined after removing OM. $\text{SSA}_{\text{occluded}}$ was the difference between the SSA of samples measured before and after removing OM. We then quantified the percent of mineral SSA covered by OM ($\% \text{SSA}_{\text{occluded}}$) (Wagai and others 2009) as:

$$\% \text{SSA}_{\text{occluded}} = \left[\frac{(\text{SSA}_{\text{total}} - \text{SSA}_{\text{untreated}})}{\text{SSA}_{\text{total}}} \right] \times 100, \quad (4)$$

where $\text{SSA}_{\text{untreated}}$ referred to mineral SSA determined before the removal of OM. Here, SSA of OM was not included because numerous measurements have shown negligible BET SSA of organic matter in soils (for example, Chiou 1990).

RESULTS

Earthworm Biomass and Species Composition

The epigeic group contributed the least to total earthworm biomass and was found across the entire transect except at 160 and 180 m (Figure 2). The endogeic group had a relatively large contribution to the earthworm biomass (Figure 2). They were not found beyond 130 m. The epi-endogeic group was among the largest contributors to biomass (Figure 2) and were found up to 190 m. The anecic group contributed to a substantial fraction of the total earthworm biomass because the individuals have the largest biomass. The distribution of anecic earthworms was variable, and they were only present at $\leq 60 \text{ m}$. Figure 2 also describes the biomass of each earthworm species by functional groups.

As shown below, endogeic species had the most profound impacts on the soil properties of interest in this study. Accordingly, we divided soil pits into two groups based on their presence: the *front group* without endogeic species consisted of pits at 150, 160, and 190 m, and the *rear group* with endogeic species was made of pits at 0, 50, and 100 m. The invasion threshold at 150 m, which was previously termed as the leading edge of the earthworm invasion (Hale and others 2005a, b), was visually determined at the location where dramatic change in leaf litter biomass occurred from patchy or no litter layer to a thick forest floor.

Carbon Concentrations and Inventories

Below the MED of 16 g cm^{-2} ($\sim 14\text{-cm}$ depth), C concentrations reached their minimum values and differences between each site were negligible. Soils in the rear group had substantially lower C concentrations than soils in the front group. In the front group, C concentrations declined to 1 % at the MED of $4\text{--}6 \text{ g cm}^{-2}$, whereas C concentrations in the rear group did so at $10\text{--}16 \text{ g cm}^{-2}$ (Figure 3a, b). The increased C % in upper A

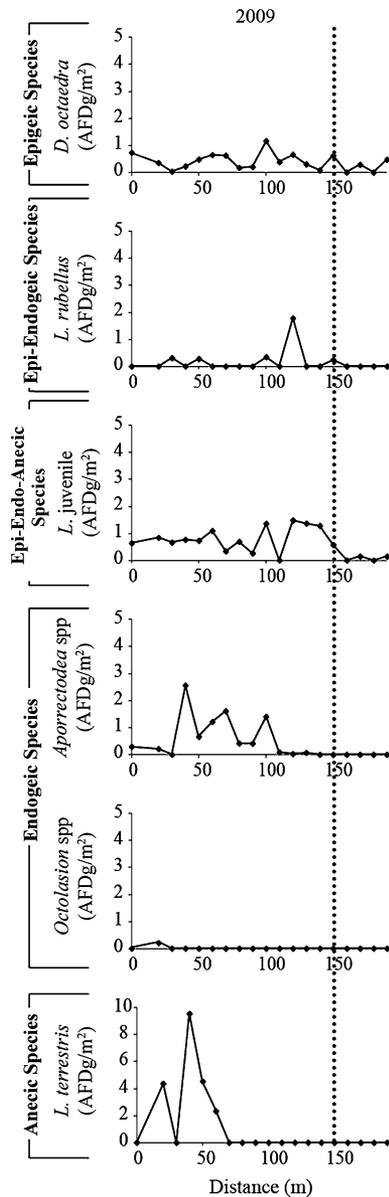


Figure 2. Earthworm biomasses along the invasion chronosequence in 2009 by species and functional grouping. The dotted line indicates the invasion threshold. In 2009, the invasion threshold was not documented. Therefore we used the long-term trend based on Hale and others (2005b) and the recent observations of the threshold in 2010 and 2011 to estimate the likely location of threshold in 2009. *L. juveniles* within the epi-endogeic group include the juveniles of the epi-endogeic *L. rubellus* and anecic *L. terrestris* that are morphologically identical and share epi-endogeic characteristics.

horizons from 190 to 150 m was juxtaposed with an abrupt reduction between 150 and 100 m presumably due to the presence of endogeic earthworms (Figure 2). Vertical homogenization of C concentrations within A horizons was evident in

soils that were inhabited by endogeic species. Additionally, we observed greater C contents at the same MEDs of the lower A and E horizons at 0 m than at other distances presumably due to the borrowing of anecic earthworms (Figure 3b).

Carbon inventories were calculated to the MEDs of 1.9 and 18.2 g cm⁻² (Figure 4). Carbon inventories decreased from 4.7 ± 0.2 kg C m⁻² in the front group to 3.2 ± 0.5 kg C m⁻² in the rear group within the MED of 1.9 g cm⁻², and the reduction in the C inventories was most evident at the endogeic forefront at 100 m. These inventories were only slightly smaller than the C inventories calculated within the MED of 18.2 g cm⁻². As such, we focus on detailing C-mineral interactions within the MED of 1.9 g cm⁻² (Figure 4).

Across the entire transect, the upper A horizons (0–1.9 g cm⁻² MED) had the largest fraction (35–69 %) of the total soil C inventories (0- to 30-cm depth). The C inventories in B horizons varied from 0.68 to 3.0 kg C m⁻². As the C concentrations and bulk densities of B horizons did not vary along the transect, the thicknesses of B horizons explained the variation in the B horizon C inventory. The four faces of excavated soil pedons (1.5 × 1.5 m²) offered large viewing areas of soil morphology and we did not find evidence of deep earthworm burrowing in the B horizons or in the lower part of the loess layers. During the excavation, we searched for earthworms in the excavated materials but failed to find them in the B horizons. Other geochemical measures covered in the next subsections provide further evidence that OM contents in the B horizons and lower part of the loess layers were not yet affected by exotic earthworms.

Changes in the upper A horizon soil C inventory along the studied transect largely arose from changes in the light density fraction. The C inventory in the light fraction above the MED of 1.9 g cm⁻² decreased from 3.4 ± 0.2 in the front group to 1.5 ± 0.7 kg C m⁻² in the rear group soils (Figure 4). To the contrary, C inventory in the heavy fraction was stable and ranged from 0.6 ± 0.2 in the front group to 1.1 ± 0.4 kg C m⁻² in the rear group.

Mineral Specific Surface Area (SSA)

Mineral SSA ranged from 0.6 to 4.2 m² g⁻¹ before OM removal (see the supplementary material Figure S-1), and the values varied from 3.6 to 34 m² g⁻¹ for the same samples once OM was removed (Figure 5). In the front group soils, individual values of SSA_{total} ranged from 5 to 37 m² g⁻¹ in

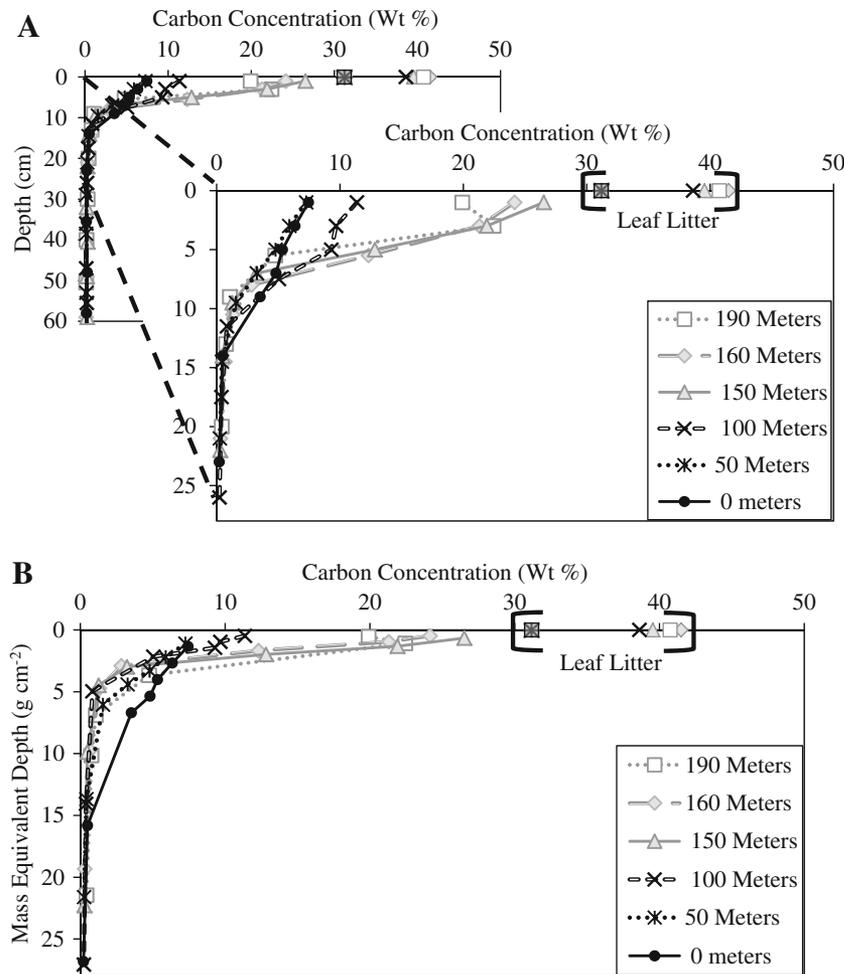


Figure 3. **A** Depth profiles of C concentrations and **B** mass equivalent depth profiles of C concentrations. Soil C inventories were estimated by integrating the C concentrations over the mass equivalent depths.

the upper A horizons. These values decreased to 6–21 m² g⁻¹ in the rear group soils. However, the deeper A horizon (MED >1.9 g cm⁻²) showed greater SSA_{total} in the rear group where soils were mixed by the endogeic species than in the front group soils. We found little change in the SSA_{total} in the loess layer below MEDs of 15 g cm⁻² (~14-cm depth).

Organic Matter Coverage of Mineral Surface

With epigeic earthworms (from 190 to 160 m), the OM-coated SSA (SSA_{occluded}) increased. However, in the areas populated with epi-endogeic earthworms (150 m), SSA_{occluded} abruptly decreased. Another step decrease occurred with endogeic earthworms (100 m). In the soils at 0 and 50 m where anecic, *L. terrestris* was present, we found a subsoil peak of SSA_{occluded}, within the deepened A horizons (Figure 6a).

The percent of mineral SSA covered by OM (%SSA_{occluded}) clearly differed between the front

and rear group soils (Figure 6b). Most (~98 %) mineral surface area was covered by OM above the MED of 1.9 g cm⁻² of the front group soils. In the rear group soils, however, less than approximately 80 % of the mineral surface area was covered by OM within the same MED.

DISCUSSION

Representativeness of the Data

In 2001, the invasion threshold was at 110 m (Hale and others 2005a). In eight years, the invasion threshold had advanced to 150 m at the averaged rate of about 5 m y⁻¹, which was comparable to the 6 to 10 m y⁻¹ that was reported for the period between 1999 and 2001 (Hale and others 2004). In addition to the consistent rates of invasion progression along the transect, the observed spatial patterns in the biomasses of different earthworm functional groups (Figure 2) agreed well with the former earthworm monitoring data at the site by Hale and others (2005b). They found that epigeic

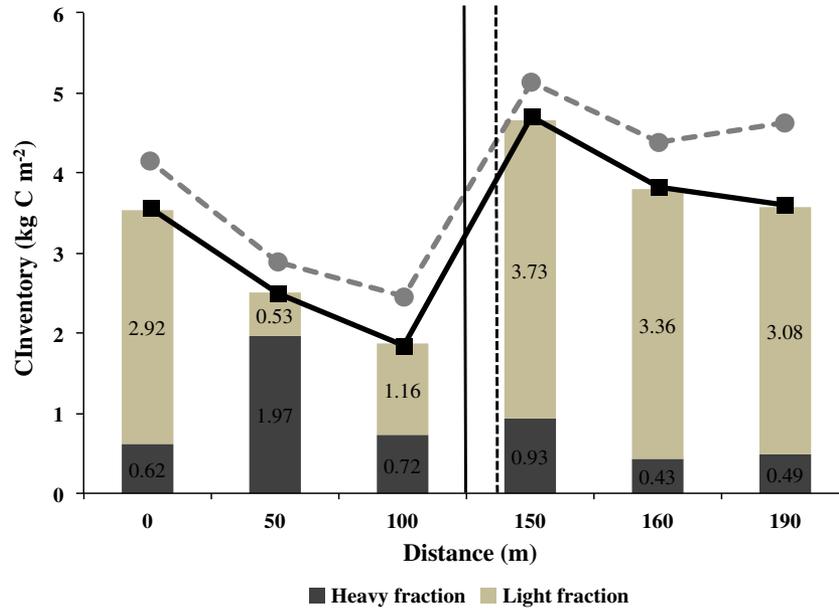


Figure 4. Soil C inventory (*solid lateral line*) in the light ($<2.0 \text{ g cm}^{-3}$) and the heavy ($>2.0 \text{ g cm}^{-3}$) fractions to the MED of 1.9 g cm^{-2} (\sim A horizon) and soil C inventory to the MED of 18.2 g cm^{-2} (*dotted lateral line*) that includes the newly created lower layer of A horizon by earthworm driven mixing and the loess layer. The *solid vertical line* represents the forefront of endogeic species that divides the studied transect into the front and rear groups, and the *dotted line* represents the earthworm invasion threshold.

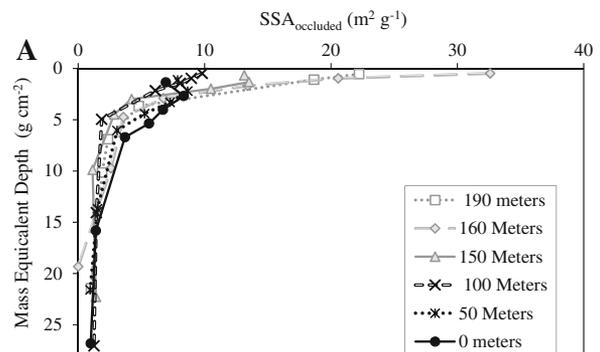
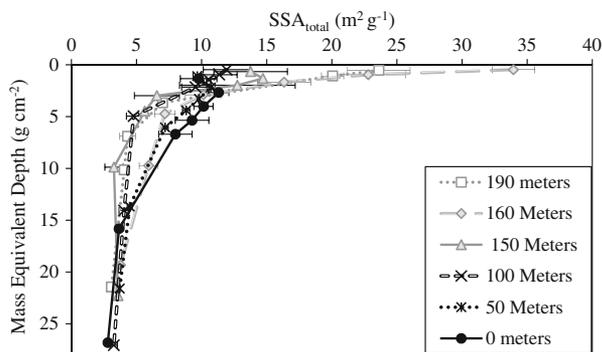


Figure 5. Mass equivalent depth profiles of $\text{SSA}_{\text{total}}$ (after the removal of organic matter). Note that the most significant changes in the values of $\text{SSA}_{\text{total}}$ between the frontal group (150, 160, and 190 m) and rear group (0, 50, and 100 m) occurred in the top 10 g cm^{-2} of MED.

earthworms were the group that first invaded along this transect, followed by endogeic, then by anecic earthworms. The existing earthworm biomass data, including the results from this study, consistently showed that the diversity of earthworm functional groups increased with the duration of earthworm invasion.

These observed rates of invasion and distributions of earthworm functional groups were mirrored by our soil morphology and C content data,

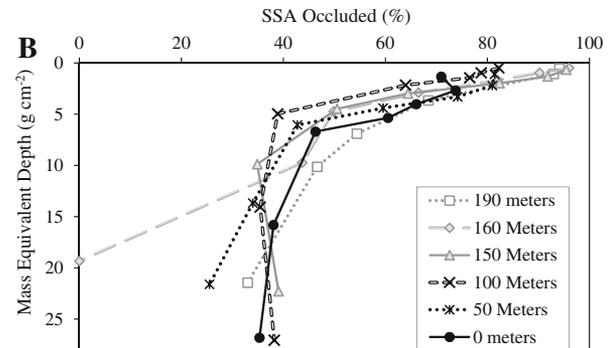


Figure 6. Mass equivalent depth profiles of **A** $\text{SSA}_{\text{occluded}}$ and **B** the percent fraction of $\text{SSA}_{\text{occluded}}$ relative to the $\text{SSA}_{\text{total}}$. Note that $\text{SSA}_{\text{occluded}}$ values changed most in the upper A horizon (that is, $<$ MED of 1.9 g cm^{-2}).

which supports our contention that six strategically placed soil pits in a design that balanced existing site knowledge with comprehensive laboratory analyses was a reasonable approach to decipher dynamic soil C responses to earthworm invasion.

Role of Exotic Earthworms in Controlling Total Mineral SSA

In all examined soils, SSA_{total} decreased with increasing soil depth in the A horizon, and this depth trend was particularly pronounced in the front group soils (Figure 5). The source of the SSA_{total} in A horizons may have been secondary phyllosilicate minerals such as illite/smectite and kaolinite; SSA_{total} and these secondary phyllosilicate minerals were similarly distributed with depth in soil profiles (see supplementary data Figure S-2. For kaolinite data, see Resner and others 2011). Kaolinite has SSA ranging from 10 to 20 $m^2 g^{-1}$, illite has SSA of 70 to 120 $m^2 g^{-1}$, and smectite has SSA of 600 to 800 $m^2 g^{-1}$, with more than 80 % of that SSA associated with internal layers of smectite (Essington 2004).

With the presence of epi-endogeic earthworms (150 and 100 m), the upper A horizon had declining SSA_{total} . This change was further accompanied by increased SSA_{total} in the lower A horizon (~ 1.9 to $6.7 g cm^{-2}$) of the soils at 0 and 50 m (Figure 5). The most straightforward explanation for this shift was vertical mixing of A horizons with underlying loess layers by epi-endogeic and endogeic species. Depth profiles of secondary phyllosilicate minerals were more homogeneous in the rear group soils (see supplementary material Figure S-2). Likewise, silt and fine sand particles from the loess layer must have been incorporated into the expanding A horizon. The loess layer, dominated by quartz and potassium plagioclases (Resner 2013), was remarkably homogeneous along the transect in its mineralogy and had a consistently low SSA of approximately $3.36 \pm 0.42 m^2 g^{-1}$ (Figure 5).

We estimated the extent to which loess layer materials were mixed into the A horizon and its impact on decreasing the averaged SSA_{total} in the upper A horizons. Our measurements of A horizon thicknesses and bulk densities (data not shown) revealed that both quantities increased with greater earthworm invasion, which had been previously shown at the study site (Hale and others 2005a). Based on the increasing mass of A horizon per ground surface area, we calculated that the exotic earthworms would incorporate approximately $0.4 kg m^{-2}$ of the loess materials

into the A horizon with each meter of advance through the forest soil.

This calculation, when combined with the SSA_{total} of the loess layer, suggested that the averaged SSA_{total} of the A horizon should have decreased by $2.7 m^2 g^{-1}$ over the length of invasion transect, which overestimated the observed reduction (Figure 7). This discrepancy may have arisen because invasive earthworms simultaneously increased pedogenic crystalline (dithionite citrate extraction) and amorphous (ammonium oxalate extraction) Fe oxides in the soils (see supplementary material Figure S-3a, b). Secondary Fe oxides extractable by dithionite citrate and ammonium oxalate are known to have high SSA (Pronk and others 2011). For instance, ferrihydrites have SSAs of 200–400 $m^2 g^{-1}$, amorphous oxides have SSAs of 305–412 $m^2 g^{-1}$, and crystalline Fe oxides have SSAs of 116–184 $m^2 g^{-1}$ (Borggaard 1982; Cornell and Schwertmann 2003). Our measurements do not inform what fraction of SSA_{total} is explained by the secondary iron oxides, but both Pronk and others (2011) and Kaiser and Guggenberger (2000) found that crystalline Fe oxides were most likely the primary sources of mineral SSA in both agricultural and forested environments.

The increase in the concentrations of the two iron oxide pools from the front to rear group soils matched the furthest advance of the endogeic earthworm at 100 m and the presence of epi-endogeic earthworm between 150 and 100 m (Figure 2). Because earthworm guts are anoxic (Drake

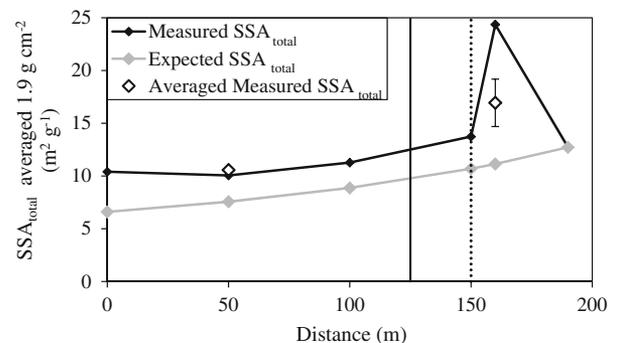


Figure 7. Measured, expected, and averaged mineral total surface area within upper A horizons. The *grey colored line* represents the expected trend of averaged SSA_{total} in A horizons (to the MED of $1.9 g cm^{-2}$) if changes in the mineral SSA_{total} in the A horizon was solely due to the mixing-driven inclusion of low SSA loess materials into the A horizon. The *dark solid line* represents actual SSA_{total} found along the transect. The *solid vertical line* divides the front and rear groups, and the *dotted vertical line* represents the invasion threshold.

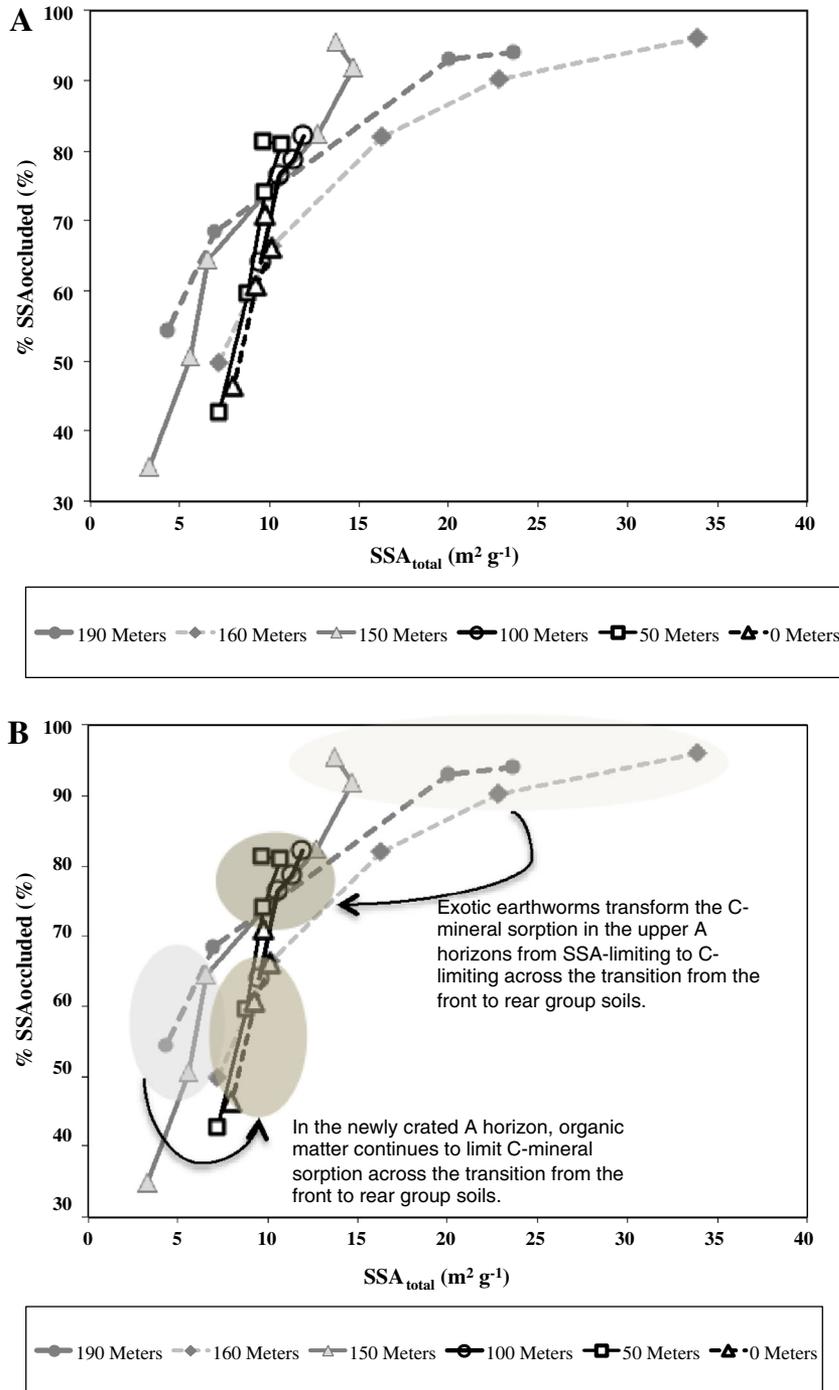


Figure 8. Relationship between the **A** %SSA_{occluded} with respect to SSA_{total} and **B** the same with annotation.

and Schramm 2006), Fe in soil materials ingested by geophagous endogeic earthworm (in particular, *Apporectodea*) may be reduced and become soluble upon passage through the gut. The dissolved Fe may later be re-oxidized and re-precipitated as Fe oxides in the aerobic environment. For example, Fe oxides increased as soil materials passed through earthworm guts (Oyedele and others 2006). Likewise, the increased SSA_{total} of the newly created

lower A horizon (~ 1.9 to 6.7 g cm⁻²) in rear group soils (Figure 5) may be explained by the physical inclusion of secondary phyllosilicate minerals into the loess layer and the biogeochemical production of secondary iron oxides.

Though not shown in Figure 5, the values of SSA_{total} of the materials in the B horizons immediately beneath the loess layer were 11.6 ± 0.9 m² g⁻¹, substantially greater than those in the loess layer.

The B horizons with peak clay contents were found to have SSA_{total} of $20.7 \pm 6.0 \text{ m}^2 \text{ g}^{-1}$. Therefore inclusion of a small amount of B horizon materials into the overlying loess layer could increase its SSA_{total} significantly. However, in agreement with the negligible presence of earthworms below the A horizon described above, there was little change in the SSA_{total} of the loess layer near the boundary to the underlying clay-rich B horizon.

Therefore, we rejected the first hypothesis that burrowing earthworms (epi-endogeic, endogeic, and anecic species) increased mineral SSA in the A horizon by incorporating clay minerals from the underlying clay-rich B horizons. Instead, earthworm bioturbation was largely limited to mixing within A horizons and between A horizons and the underlying loess at the study site.

Limiting Factors for OM Sorption on Mineral Surface

We hypothesized that greater mixing by earthworms would increase the size and fraction of OM-occluded mineral surface area by enhancing physical contact between OM and mineral surfaces. Our results were not consistent with this hypothesis. The soils inhabited only by epigeic earthworm species have not only larger $SSA_{occluded}$ but also generally higher percentages of SSA_{total} covered with OM in upper A horizons (Figure 6a, b). With the arrival of epi-endogeic and endogeic earthworms, less mineral surface area was present in upper A horizons (discussed above) and smaller fractions of the surface area were covered with OM. It is also notable that the upper A horizon of the soil at 150 meter represented the transition in $SSA_{occluded}$ from the front to rear groups but clearly belonged to the front group in terms of $\%SSA_{occluded}$.

It appears that OM-mineral sorption was limited by available mineral surface area prior to the arrival of endogeic earthworms (Figure 8). Within the MED of 1.9 g cm^{-2} , no less than 97 % of the total mineral surface area, over the range from 13 to $35 \text{ m}^2 \text{ g}^{-1}$, was consistently covered by OM (Figure 8b). It is reasonable to assume that the remaining approximately 3 % of the OM-free mineral surface area originated from primary minerals like quartz and plagioclase which are non-reactive for the sorption of OM. All available mineral surface was occluded by OM where the highest contents of light-fraction carbon were also observed.

With the arrival of endogeic earthworms, however, OM-sorption on mineral surface in the upper A

horizon appears to become limited by mineral-free or light fraction OM (Figure 8b). The inventory of light fraction C in the upper A horizon was reduced by half from $3.4 \pm 0.2 \text{ kg C m}^{-2}$ in the front group soils to $1.5 \pm 0.7 \text{ kg C m}^{-2}$ in the rear group soils (Figure 4) as endogeic earthworms consumed and vertically rearranged soil OM. To examine mineral availability for OM sorption, we calculated the inventory of total mineral surface area within the same upper A horizon where MED was 0 to 1.9 g cm^{-2} (Figure 9a). There was $0.32 \pm 0.046 \times 10^6 \text{ m}^2$ of SSA_{total} in the upper A horizon per ground surface area of 1 m^2 in the front group. This value decreased to $(0.19 \pm 0.044) \times 10^6 \text{ m}^2$ in the rear group, a reduction by 40 % (Figure 9a). A still smaller fraction of the mineral surface area was covered by OM in the rear group soils than in the front group soils. Such dramatic reductions in SSA_{total} were still less than the reduction of light fraction C (Figure 4). Such disproportionate reductions in mineral surface area and mineral-free OM

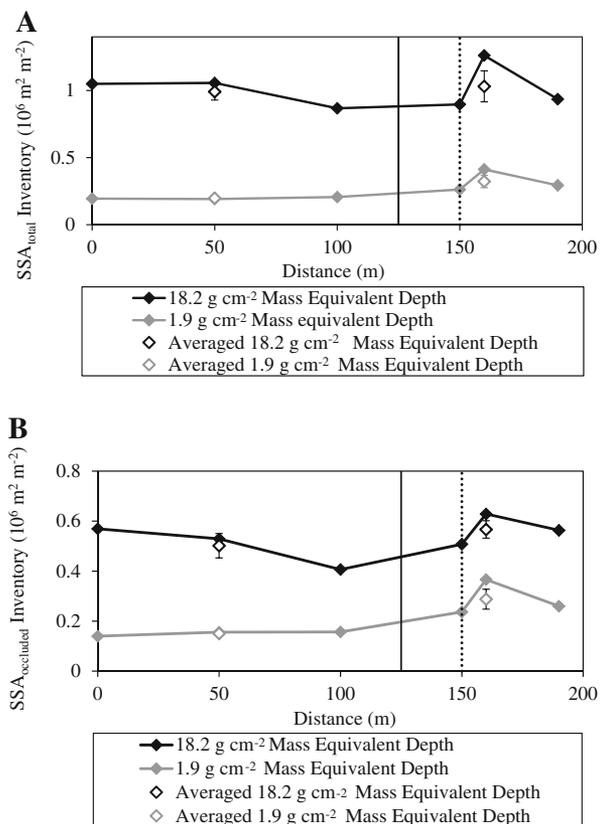


Figure 9. A-horizon inventories of **A** SSA_{total} and **B** $SSA_{occluded}$. Changes in soil bulk densities and A horizon thicknesses were considered in the calculation of inventories. The *solid line* separates the front and rear groups, and *dotted line* shows the earthworm invasion threshold.

may have caused a shift from mineral-limitation to OM-limitation in controlling OM-sorption on minerals with the invasion of endogeic earthworms.

Regarding this transition from mineral-limitation to OM-limitation, the upper A horizon at 150 meter offers an opportunity to examine a transitional state. Here, epi-endogeic earthworms had incorporated loess materials into the A horizon and reduced SSA_{total} in the upper A horizon (Figure 5). Such a reduction in SSA_{total} occurred, however, while the C content in the upper A horizon was reaching the maximum values observed over the entire transect (Figure 3b). Here, epi-endogeic mixing of litter layer into mineral soil had enriched carbon content in the upper A horizon, but the accelerated endogeic consumption of the organic matter in the upper A horizon had not yet occurred. The combined effect was that OM-mineral sorption in the upper A horizon continued to be limited by mineral surface area.

For the lower A horizons that were newly created by mixing of A horizons and loess layers, the slight increase in the SSA_{total} in the rear group soils occurred due to mixing of secondary phyllosilicate clay mineral and iron oxides from the pre-existing A horizons into the loess layer. Such increases in SSA_{total} , however, did not result in similar increases in $SSA_{occluded}$ (Figure 6a, b). We concluded that, as the upper portion of the loess layer was mixed into lower A horizons by earthworms, this zone continued to be OM-limiting in terms of OM-sorption on minerals.

Therefore, within the A horizons, there were contrasting changes in factors that limited C-mineral sorption along the earthworm invasion transect. The A horizon thickness varied along the transect. If we had considered the A horizon as one entity affected by earthworm activity, which has been typical of biogeochemical studies of invasive earthworms, we would not have identified changes in mechanisms that controlled C-mineral sorption. It should be, however, noted that the contribution of newly created lower A horizon in the soil C inventory was negligible (Figure 4) relative to the upper A horizon.

Organic Matter Sorption on Mineral Surface and Soil Carbon Inventory

Despite reductions in A horizon inventories of C across the transition from the front to rear group soils, the inventories of the heavy fraction C remained indistinguishable across the transition (Figure 4). Although a limited number of soil sampling pits prevents generalization, it is notable

that the heavy fraction C inventory was particularly high at 50 m where the presence of endogeic and anecic species was strongest (Figure 2). This trend, however, was not consistent with the observed $SSA_{occluded}$. The inventories of $SSA_{occluded}$, within the MED of 1.9 g cm^{-2} , decreased from $0.29 \pm 0.04 \times 10^6 \text{ m}^2 \text{ m}^{-2}$ in the front group to $0.15 \pm 0.005 \times 10^6 \text{ m}^2 \text{ m}^{-2}$ in the rear group (Figure 9a, b). Two potential mechanisms may have contributed to the discrepancy. Coverage of mineral surface with OM was found to be patchy rather than continuous (Kaiser and Guggenberger 2003; Ransom and others 1998), and multi-layered (Kleber and others 2007) rather than mono-layered (Mayer 1999). As endogeic earthworms affect OM-mineral sorption by mixing soils and ingest OM and minerals, OM sorption on mineral surface may have become more patchy resulting in reduced $\%SSA_{occluded}$.

Second, the persistent heavy fraction OM may be due to increased aggregation in soils inhabited by endogeic species. Along the studied transect, the pre-endogeic soils had A-horizon materials that were largely structureless and free of aggregates. However, A-horizon soils with endogeic species had strong medium size granular structure, which may have contributed to physically occluding OM and thus maintaining heavy fraction carbon. The way that mineral-occlusion of C responded to earthworm bioturbation may thus be distinct from that of mineral-C sorption. Although the decreases in mineral SSA and OM negatively affected C sorption onto mineral surface in this study, our field observation revealed a greater degree of mineral aggregation. Therefore, impacts of exotic earthworms on mineral-associated soil C inventory may not only be dependent upon absolute and relative abundances of soil OM and mineral surface area and sorption mechanisms but also be subject to somewhat independent formation of aggregates affected by exotic earthworms.

Studies on soil C inventories, conducted in earthworm infected deciduous forests in Minnesota and New York, USA, offered compounding results regarding the positive or negative response of soil C inventory to exotic earthworms (for example, Alban and Berry 1994; Burtelow and others 1998; Bohlen and others 2004a and b; Wironen and Moore 2006). These conflicting outcomes may reflect differences among stages of earthworm invasion and also the specifics of earthworm-derived OM-mineral interactions within a given ecosystem that are constrained by soil properties. Therefore, we feel that studies of the specifics of C-stabilization on soils may provide valuable insight to

reconcile why findings have varied among earthworm-invaded sites.

CONCLUSION

We rejected our hypotheses that earthworms would increase mineral SSA of A horizon materials by upwardly moving the clay rich B horizon materials into A horizons and would enhance OM sorption on mineral surfaces by mixing them. Both endogeic and anecic earthworms did not burrow below the loess layer at our site. Had the A horizon directly overlain a less dense clay rich B horizon, earthworm driven soil mixing may have increased SSA_{total} in the A horizon with a potential to sorb more OM. This scenario suggests that the effects of invasive earthworms on C-mineral sorption and thus soil C storage could be further determined by initial soil properties such as horizonation, mineralogy, and texture and the behavioral responses of exotic earthworms to the soil properties.

Our study highlights that the limiting factor on OM-mineral sorption in the upper A horizons shifted from available mineral surface area before endogeic invasion to available OM after endogeic invasion. In other words, exotic endogeic earthworms deplete the available OM pool at a rate faster than sorption of OM on new mineral surface. At our study site, earthworm driven mixing thickened A horizon. However, as our analysis showed, C-mineral sorption in this newly created zone remained OM-limiting and was far from negating the loss of C observed in the upper A horizon. Whether there exist systems where new OM-mineral sorption balances or outweighs OM-consumption is an intriguing question that remains to be tested. This shift in OM-mineral interactions occurred within 10 years of the arrival of endogeic earthworms, illustrating the efficiency with which exotic earthworms affect the key ecosystem process in formerly earthworm-free glaciated forests. We refuted our original hypotheses but determined some underlying mechanisms, which shows a strong potential for applying this approach to other soil systems influenced by bioturbators.

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REFERENCES

- Adams MB, Loughry LH, Plaughter LP. 2004. Experimental Forests and Ranges of the USDA Forest Service. Newtown Square (PA): US Department of Agriculture.
- Alban DH, Berry EC. 1994. Effects of earthworm invasion on morphology, carbon and nitrogen of a forest soil. *Appl Soil Ecol* 1:243–9.
- Baldock JA, Skjemstad JO. 2000. Role of the soil matrix and minerals in protecting natural organic materials against biological attack. *Org Geochem* 31(7):697–710.
- Bohlen PJ, Pelletier D, Groffman PM, Fahey TJ, Fisk MC. 2004a. Ecosystem consequences of exotic earthworm invasion of north temperate forests. *Ecosystems* 7:1–12.
- Bohlen PJ, Pelletier DM, Groffman PM, Fahey TJ, Fisk MC. 2004b. Influence of earthworm invasion on redistribution and retention of soil carbon and nitrogen in northern temperate forests. *Ecosystems* 7:13–27.
- Bohlen PJ, Scheu S, Hale CM, McLean MA, Migge S, Groffman PM, Parkinson D. 2004c. Non-native invasive earthworms as agents of change in northern temperate forests. *Front Ecol Environ* 2(8):427–35.
- Borggaard OK. 1982. The influence of iron oxides on the surface area of soil. *J Soil Sci* 33:443–9.
- Burtelow AE, Bohlen PJ, Groffman PM. 1998. Influence of exotic earthworm invasion on soil organic matter, microbial biomass and denitrification potential in forest soils of the northeastern United States. *Appl Soil Ecol* 9:197–202.
- Chiou CT. 1990. The surface area of soil organic matter. *Environ Sci Technol* 24:1164–6.
- Cornell RM, Schwertmann U. 2003. The iron oxides: structure, properties, reactions, occurrences and uses. Weinheim (DE): Wiley. p 664.
- Crooks J. 2002. Characterizing ecosystem-level consequences of biological invasions: the role of ecosystem engineers. *Oikos* 97:153–66.
- Drake HL, Schramm HMA. 2006. Earthworm gut microbial biomes: their importance to soil microorganisms, denitrification, and the terrestrial production of the greenhouse gas N_2O . *Soil Biol* 6:65–87.
- Eberl DD. 2003. User's guide to Rockjock: a program for determining quantitative mineralogy from powder X-ray diffraction data. US Geological Survey Open-File Report: 03-78. p. 46.
- Essington ME. 2004. Soil and water chemistry: an integrative approach. Boca Raton (FL): CRC Press. p 534.
- Eusterhues K, Rumpel C, Kögel-Knabner I. 2005. Organo-mineral associations in sandy acid forest soils: importance of specific surface area, iron oxides and micropores. *Eur J Soil Sci* 56:753–63.
- Frelich LE, Hale CM, Scheu S, Holdsworth AR, Henegham L, Bohlen PJ, Reich PB. 2006. Earthworm invasion into previously earthworm-free temperate and boreal forests. *Biol Invasions* 8:1235–45.
- Gundale MJ. 2002. The influence of exotic earthworms on soil organic horizon and the rare fern *Botrychium mormo*. *Conserv Biol* 16:1555–73.

- Hale CM, Frelich LE, Reich PB. 2004. Allometric equations for estimation of ash-free dry mass from length measurements for selected European earthworm species (Lumbricidae) in the western Great Lakes region. *Am Midl Nat J* 151(1):179–85.
- Hale CM, Frelich LE, Reich PB, Pastor J. 2005a. Effects of European earthworm invasion on soil characteristics in northern hardwood forests of Minnesota, USA. *Ecosystems* 8:911–27.
- Hale CM, Frelich LE, Reich PB. 2005b. Exotic European earthworm invasion dynamics in northern hardwood forests of Minnesota, USA. *Ecol Appl* 15:848–60.
- Hale CM, Frelich LE, Reich PB. 2006. Changes in cold-temperate hardwood forest understory plant communities in response to invasion by European earthworms. *Ecology* 87:1637–49.
- Hale CM. 2007. Earthworms of the Great Lakes. Duluth (MN): Kollath and Stensaas Publishing. p 36.
- Hale CM, Frelich LE, Reich PB, Pastor J. 2008. Exotic earthworm effects on hardwood forest floor, nutrient availability and native plants: a mesocosm study. *Oecologia* 155:509–18.
- Hendrix PF, Callahan MA, Drake JM, Huang CY, James SW, Snyder BA, Zhang W. 2008. The global problem of introduced earthworms. *Ann Rev Ecol Evol Syst* 39:593–613.
- Hendrix PF, Bohlen PJ. 2002. Exotic earthworm invasions in North America: ecological and policy implications. *BioScience* 52(9):801–11.
- Holmgren GGS. 1967. A rapid citrate-dithionate extractable iron procedure. *Soil Sci Soc Am Proc* 31:210–11.
- Kaiser K, Guggenberger G. 2000. The role of DOM sorption to mineral surfaces in the preservation of organic matter in soils. *Org Geochem* 31:711–25.
- Kaiser K, Guggenberger G. 2003. Mineral surfaces and soil organic matter. *Eur J Soil Sci* 54:219–36.
- Kaiser K, Guggenberger G. 2007. Sorptive stabilization of organic matter by microporous goethite: sorption into small pores vs. surface complexation. *Eur J Soil Sci* 58:45–59.
- Kleber M, Sollins P, Sutton R. 2007. A conceptual model of organo-mineral interactions in soils: self-assembly of organic molecular fragments into zonal structures on mineral surfaces. *Biogeochemistry* 85:9–24.
- Lawrence AP, Bowers MA. 2002. A test of the 'hot' mustard extraction method of sampling earthworms. *Soil Biol Biochem* 34:549–52.
- Mayer LM. 1999. Extent of coverage of interal surfaces by organic matter in marine sediments. *Geochim Cosmochim Acta* 63(2):207–15.
- McKeague JA, Day JH. 1966. Dithionite and oxalate-extractable Fe and Al as aids in differentiating various classes of soils. *Can J Soil Sci* 46:13–22.
- Minnesota State Climatology Office. 2003. Minnesota climatology working group website <http://climate.umn.edu>, Minnesota Department of Natural Resources and the University of Minnesota, Department of Soil, Water and Climate, St. Paul, MN 55108.
- Oyedele DJ, Schjønning P, Amusan AA. 2006. Physiochemical properties of earthworm casts and ungested parent soil from selected sites in southwestern Nigeria. *Ecol Eng* 28:106–13.
- Pronk GJ, Heister K, Kögel-Knabner I. 2011. Iron oxides as major available interface component in loamy arable topsoils. *Soil Sci Soc Am J* 75:2158–68.
- Ransom B, Kim D, Kastner M, Wainwright S. 1998. Organic matter preservation on continental slopes: importance of mineralogy and surface area. *Geochim Cosmochim Acta* 62:1329–45.
- Remusat L, Hatton Pierre-Joseph, Nico Peter S, Zeller Bernd, Kleber Markus, Derrien Delphine. 2012. NanoSIMS study of organic matter associated with soil aggregates: advantages, limitations, and combination with STXM. *Environ Sci Technol* 46:3943–9.
- Resner KE. 2013. Impacts of earthworm bioturbation on elemental cycles in soils: An application of a geochemical mass balance to an earthworm invasion chronosequence in a sugar maple forest in Northern Minnesota. MS thesis. University of Minnesota. p 182.
- Resner KE, Yoo K, Hale C, Aufdenkampe A, Blum A, Sebestyen S. 2011. Elemental and mineralogical changes in soil due to bioturbation along an earthworm invasion chronosequence in northern Minnesota. *Appl Geochem* 26:S127–31.
- Reynolds JW. 1977. The earthworms (Lumbricidae and Sparganophilidae) of Ontario. Toronto (ON): Royal Ontario Museum Miscellaneous Publication.
- Scheu S, Parkinson D. 1994a. Effects of earthworms on nutrient dynamics, carbon turnover and microorganisms in soil from cool temperate forests of the Canadian Rocky Mountains: laboratory studies. *Appl Soil Ecol* 1:113–25.
- Scheu S, Parkinson D. 1994b. Effects of invasion of an aspen forest (Canada) by *Dendrobaena octaedra* (Lumbricidae) on plant growth. *Ecology* 75:2348–61.
- Schmidt MWI, Torn MS, Abiven S, Dittmar T, Guggenberger G, Janssens IA, Kleber M, Kögel-Knabner I, Lehmann J, Manning DAC, Nannipieri P, Rasse DP, Weiner S, Trumbore SE. 2011. Persistence of soil organic matter as an ecosystem property. *Nature* 478:49–56.
- Schwert, DP. 1990. Oligochaeta: Lumbricidae. Dindal DL, editor. *Soil biology guide*. New York: John Wiley and Sons. p.341–356.
- Sollins P, Kramer M, Swantson C, Lajtha K, Filley T, Aufdenkampe A, Wagai R, Bowden R. 2009. Sequential density fractionation across soils of contrasting mineralogy: evidence for both microbial- and mineral-controlled soil organic matter stabilization. *Biogeochemistry* 96:209–31.
- Torn MS, Trumbore SE, Chadwick OA, Vitousek PM, Hendricks MD. 1997. Mineral control of soil organic carbon storage and turnover. *Nature* 389:170–3.
- Torresan M. 1987. The use of sodium polytungstate in heavy mineral separations. US Geological Survey Open-File Report. pp 87–590
- von Lutzow M, Kögel-Knabner I, Ekschmitt K, Matzner E, Guggenberger G, Marschner B, Flessa H. 2006. Stabilization of organic matter in temperate soils: mechanisms and their relevance under different soil conditions: a review. *Eur J Soil Sci* 57:426–45.
- Wagai R, Mayer LM, Kitayama K. 2009. Extent and nature of organic coverage of soil mineral surfaces assessed by a gas sorption approach. *Geoderma* 149:152–60.
- Webb PA, Orr C. 1997. Analytical methods in fine particle technology. Norcross (GA): Micromeritics Instrument Corp. p 301.
- Wironen M, Moore TR. 2006. Exotic earthworm invasion increases soil carbon and nitrogen in an old-growth forest in southern Quebec. *Can J For Res* 36:845–54.